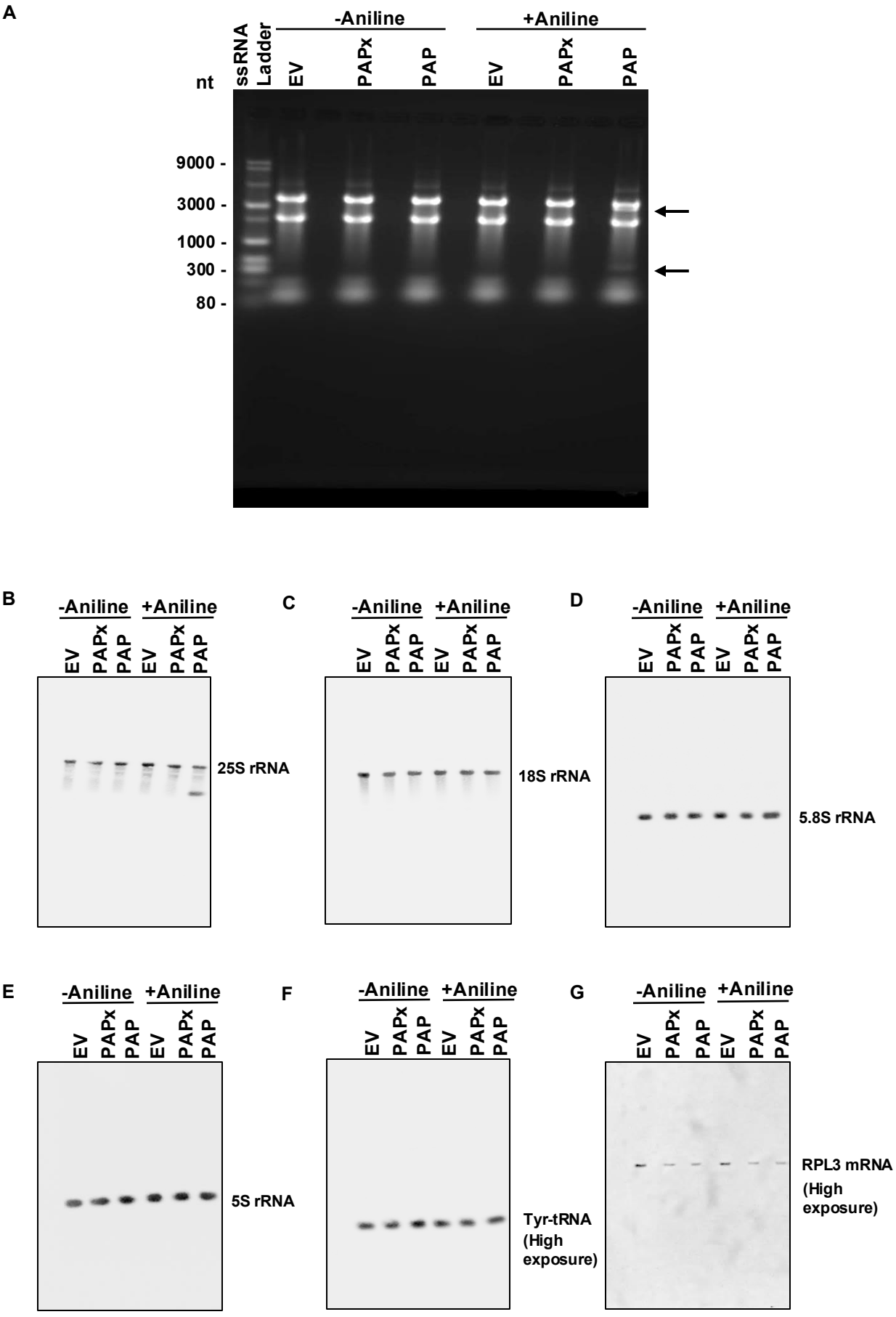

Supplemental Figure S2. Detection of RNA using SYBR gold-stained agarose gel and northern blotting. (A) Total RNA was isolated from WT BY4741 yeast strain transformed with EV, PAPx and PAP cDNAs following 6 hours of induction in 2% galactose. RNA samples (2.5 µg), treated with or without aniline, were separated on a 1.5% formamide agarose gel. The gel was stained with 10 µL of 10000X SYBR gold stain in 50 mL of 1X TAE buffer for 20 minutes and destained for 10 minutes in dH_2O. A single-stranded RNA ladder is indicated with band sizes. An image was acquired using a UV transilluminator. Black arrows indicate RNA fragments consistent with cleavage at A3027 of 25S rRNA. (B-G) 2.5 µg of RNA samples treated with or without aniline, were separated on a 1.5% agarose gel and transferred to a nylon membrane. The membrane was probed with the 5' biotinylated probes specific for (B) 25S rRNA (C) 18S rRNA (D) 5.8S rRNA (E) 5S rRNA (F) tyrosyl-tRNA (G) ribosomal protein L3 mRNA (RPL3). Bands were visualized with a chemiluminescence detection system. The sequences of the 5' biotinylated probes are listed in Supplemental Table S3.