

Supplementary materials and methods

Determination of the structure of N7-guanosine using NMR methods

Structures of N7-guanosine (7G) were confirmed by 1D ^1H and ^{13}C NMR experiments and chemical shifts for 7G were in good agreement with literature data (Figure S8 and S9) (Wawrzyniak et al. 2020). Additionally, assignment of ^1H , ^{13}C and ^{15}N resonances was confirmed by the analysis of 2D spectra: ^1H - ^1H correlation spectroscopy (COSY) as well as ^1H - ^{13}C gradient heteronuclear single quantum coherence (gHSQC) and gradient heteronuclear multiple bond coherence (gHMBC) spectroscopies and compared with the data obtained for guanosine (Tables S4 and S5, Figure S10-S14).

As previously reported, 9 $\rightarrow$ 7 isomerization of guanosine caused significant differences in ^{13}C and ^1H resonances, especially for carbons of base moiety, *i.e.* C4, C5, and C8, as well as for protons H8, H1' and NH_2 (Table S4 and S5) (Baranowski et al. 2022). Conspicuous upfield shift by 10 ppm for C5 and downfield shift by 8 and 6 ppm for C4 and C8, respectively, were observed, which was a characteristic pattern of changes of ^{13}C resonances for N7 isomer of guanosine compared with N9 isomer. Smaller upfield shifts *i.e.* 2-3 ppms were also observed for C2 and C6. For ^1H resonances, signals of H8 and H1' underwent a downfield shift by 0.2-0.3 ppm. On the other hand, the signal of NH_2 showed upfield shifts by 0.3 ppm.

The characteristic heteronuclear long range correlations *i.e.* C5-H1' and C8-H1' observed on ^1H - ^{13}C gHMBC spectrum were crucial for unambiguous determination of D-ribofuranosyl substitution at N7 position of guanosine (Figure S12).

Calculation of free energies of the N7-guanosine and guanosine tautomers

Density functional theory (DFT) calculations at B3LYP/6-311G(d) level of theory were performed on N7-methylated guanine (m^7Gua) representing the 7G-tautomers obtained from the QM optimized structures (at HF/6-311G(d) level of theory) of the 6 different keto-enol tautomers (replacing the ribose sugar with a methyl group) to obtain the free energies of these tautomers (m^7Gua tautomers)

(Figure S4b). Similar calculations were performed on N9-methylated guanine ($m^9\text{Gua}$) tautomers representing guanosine (G)-tautomeric forms (Figure S4a). The $m^9\text{Gua}$ tautomers were prepared using MOLDEN (Schaftenaar and Noordik 2000) and subjected to optimization at HF/6-311G(d) theory and the energy calculations (at B3LYP/6-311G(d) level of theory) were performed using the optimized structures. The free energy values were estimated both in the gas phase and the solvent phase (Figure S5). For the calculations in the solvent phase water environment by the polarizable continuum model (PCM) was considered.

Reoptimization of glycosidic torsion parameters for 7G-N1H

Four initial geometries (conformational schemes SC1-4) of the 7G-N1H residue were prepared following Yildirim et al. (Yildirim et al. 2010) and subjected to geometry optimization followed by a gas phase potential energy surface (PES) scan executed around the glycosidic torsion angle ($\chi+180^\circ$ i.e. O4'-C1'-N7-C8) with an increase in the value of ($\chi+180^\circ$) by 5° resulting in 72 conformations (for each scheme) following the protocol described in Yildirim et al. (Yildirim et al. 2010). The QM energies of each of the 72 conformations (for each scheme) were calculated using the MP2/6-31G* level of theory (Figure S15). The quantum mechanically (at HF/6-31G* level of theory) optimized lowest energy conformer was used for the calculation of partial atomic charges using RESP fitting using the R.E.D. version III.52 perl program Vanqualef et al, (Vanqualef et al. 2011). Next, the molecular mechanical (MM) energy minimization was performed following the protocol described earlier (Dutta et al. 2023) for the development of glycosidic torsion parameters within the AMBER FF99 force field (Wang et al. 2000). We also followed a similar protocol for fitting of glycosidic torsion potential as described earlier (Dutta et al. 2023). In this approach, the potential energy due to the glycosidic torsion (χ) is obtained from the differences between the QM energies (E_{QM}) and the MM energies (E_{MM}) i.e. from the equation: $E_\chi = E_{\text{QM}} - E_{\text{MM}}$. $4 \times 72 = 288$ E_χ values calculated for the 288 conformations using the aforementioned equation, were fitted to the Fourier series shown in the

following equation: $E(\phi_1, \phi_2) = \sum_{\phi=1}^4 V_{n1}[1 + \cos(n\phi_1)] + V_{n2}[1 + \cos(n\phi_2)]$ (here, V_{n1} , V_{n2} , ϕ_1 and ϕ_2 are varied). ϕ_1 and ϕ_2 represent the (O4'-C1'-N7-C8) and (C2'-C1'-N7-C8) torsion angles respectively and V_{n1} and V_{n2} represent the potential energy barriers around these torsion angles. A separate fitting was performed for calculating V_{n1} and V_{n2} . The QM and MM energy profiles (with χ set to zero and with the newly derived χ_{KOL0} parameters) for 7G-N1H are shown in Figure S16.

For 7G-N3H a similar protocol as described for 7G-N1H was followed for geometry optimization, PES scan, QM energy calculations was followed and the lowest energy conformer was subjected to further optimization, followed by RESP fitting. The lowest energy conformer of 7G-N1H (based on the QM energy calculations) was modified in MOLDEN to prepare the 7G-N1H,N2-imino tautomer, followed by geometry optimization at HF/6-31G* level of theory and subsequent calculation of partial atomic charges using RESP fitting. As mentioned in the main text, for the 7G-N3H and 7G-N1H,N2-imino tautomers, the glycosidic torsion parameters developed for the 7G-N1H tautomer were applied during MD simulations of duplexes.

Calculation of hybridization energies of duplexes using MM/3D-RISM

As mentioned in the main text, the duplex hybridization energies (binding free energies) for the duplexes in this study were estimated using the molecular mechanics/three-dimensional reference interaction site model (MM/3D-RISM) approach and only the internal seven base pairs were considered during the calculations (i.e. prior to the calculations, both the terminal base pairs were removed). In this approach, 3D-RISM (three-dimensional reference interaction site model) is utilized for the description of the three-dimensional map of solvent density distribution around the solute molecule (Kovalenko and Hirata, 2003; Luchko et al. 2010). Here, in the 3D space (r), the solvent structure is represented using the probability density $\rho_\gamma g_\gamma(r)$ (which approaches the solvent bulk density ρ_γ with $g_\gamma(r) \rightarrow 1$, where $g_\gamma(r)$ is the 3D solvent site distribution function) of finding the solvent sites (γ) around the solute (Luchko et al. 2010). Additionally, we have also utilized the 3D version of

Kovalenko-Hirata closure (3D-KH closure) (Genheden et al. 2010) molecular solvation theory during the MM/3D-RISM calculations. The 3D-RISM-KH approach has been previously utilized for the calculation of binding free energies for short DNA oligonucleotides by Yesudas et al, (Yesudas et al. 2015) and was observed to produce good agreement with experimental results. This approach has also been used for the dsRNA oligonucleotides by Yildirim et al, (Yildirim et al. 2023). In our calculations we used a similar script as provided in Yildirim et al, (Yildirim et al. 2023) and we used the parameters as shown below: `/&rism closure="kh", rism_verbose=0, polardecomp=0, thermo="gf"/`.

Supporting Tables

Table S1. (A) Calculated properties of the duplexes containing 7G-A base pair from MD simulations

	5'G7GC/3'CAG duplex		5'C7GG/3'GAC duplex	
	G7GC (7G(N1H)-A) (GM1)	G7GC (7G(N1H,N3H,N2-imino)-A) (GM3)	C7GG (7G(N1H)-A) (GM2)	C7GG (7G(N1H,N3H,N2-imino)-A) (GM3)
RMSD (Å)	1.5 (0.4)	1.3 (0.3)	1.5 (0.4)	1.4 (0.3)
C1'-C1' distance (Å)	10.9 (0.3)	10.8 (0.2)	10.6 (0.3)	10.7 (0.2)
7G5 (C1'-C1'-N7) (°)	82.7 (4.4)	85.6 (3.3)	30.7 (3.4)	85.2 (11.9)
A14 (C1'-C1'-N9) (°)	65.1 (7.8)	64.1 (3.9)	90.3 (5.0)	65.9 (7.8)
NOE distances (Å)				
A14-H2...7G-H1 Population with distance <3 Å (strong NOE)	2.5 (0.7) 94%	2.8 (0.3) 83%	5.4 (0.2) 0.0%	2.8 (0.2) 83%
A14-H2 ...C6-H1' Population with distance <4.5 Å (medium NOE)	4.3 (0.5) 66%	4.2 (0.5) 71%	NA	NA
A14-H2 ...G6-H1' Population with distance <4.5 Å (medium NOE)	NA	NA	6.9 (0.6) 0.01%	4.2 (0.4) 72%
A14-H2 ...G6-H1 Population with distance <6 Å (Weak NOE)	NA	NA	4.3 (0.6) 99%	4.6 (0.4) 99%

A14-H2 ...C15-H1' Population with distance <4.5 Å (medium NOE)	4.3 (0.6) 64%	4.2 (0.6) 70%	NA	NA
A14-H2 ...G15-H1' Population with distance <4.5 Å (medium NOE)	NA	NA	6.2 (0.5) 0.08%	4.3 (0.6) 65%
A14-H2 ...G15-H1 Population with distance <6 Å (Weak NOE)	NA	NA	5.0 (0.6) 94%	5.3 (0.4) 95%
A14-H2 ... G13-H1 Population with distance <4.5 Å (medium NOE)	3.7 (0.5) 96%	3.8 (0.4) 95%	NA	NA
A14-H2 ... C13-H41 Population with distance <6 Å (weak NOE)	NA	NA	4.7 (0.5) 99%	6.7 (0.5) 6%
The values within parentheses correspond to standard deviations.				

(B) Calculated properties of the duplexes containing 7G-G base pair from MD simulations

Duplexes	5'G7GC/3'CGG duplex			5'C7GG/3'GGC duplex		
	G7GC (7G(N1H)-G) (GM1)	G7GC (7G(N1H)-G) (GM2)	G7GC (7G(N3H)- G) (GM3)	C7GG (7G(N1H)-G) (GM1)	C7GG (7G(N1H)-G) (GM2)	C7GG (7G(N3H)-G) (GM3)
RMSD (Å)	1.7 (0.3)	1.6 (0.4)	1.9 (0.4)	1.9 (0.5)	1.4 (0.3)	1.5 (0.3)
C1'-C1' distance (Å)	11.1 (0.3)	10.6 (0.3)	11.1 (0.2)	11.2 (0.3)	10.4 (0.3)	11.1 (0.2)
7G5 (C1'-C1'-N7) (°)	90.3 (3.8)	29.8 (4.3)	84.2 (3.3)	90.4 (3.8)	30.7 (3.7)	84.4 (3.4)
G14 (C1'-C1'-N9) (°)	46.7 (4.3)	98.9 (4.6)	60.5 (3.3)	43.8 (4.1)	101.3 (4.1)	61.0 (3.4)
NOE distances (Å)						

G14_H1...H1_G6 Population with distance <6 Å (weak NOE)	NA	NA	NA	4.9 (0.4) 99%	5.6 (0.5) 81%	4.5 (0.4) 100%
G14_H1...H8_G6 Population with distance <6 Å (weak NOE)	NA	NA	NA	4.6 (0.5) 99%	8.7 (0.6) 0.01%	5.5 (0.5) 88%
G14_H1...H8_G15 Population with distance <6 Å (weak NOE)	NA	NA	NA	4.9 (0.4) 99%	6.6 (0.4) 8%	5.0 (0.4) 99%
The values within parentheses correspond to standard deviations.						

Table S2. (A) Energetics of the duplexes with 7G-A pair

Duplexes	Duplex - GGC (G-A) Anti-Syn Syn-Anti Anti-Anti (Geometry-1 95%) Anti-Anti (Geometry-2 3%)	Duplex - G7GC (7G(N1H)-A) (GM1)	Duplex - G7GC (7G(N1H,N3H,N2- imino)-A) (GM3)	Duplex - CGG (G-A) Anti-Syn Syn-Anti (200-360ns) Anti-Anti (Geometry-1 81%) Anti-Anti (Geometry-2 13%)	Duplex - C7GG (7G(N1H)-A) (GM2)	Duplex - C7GG (7G(N1H,N3H,N2- imino)-A) (GM3)
Hybridization energies (ΔG) and changes in hybridization energies ($\Delta\Delta G$) upon modification						
ΔG (MM/3D- RISM) (kcal/mol)	-33.2 (4.6) ^a -27.0 (4.3) -27.1 (4.2) -30.1 (4.7)	-30.5 (4.5)	-36.8 (4.4)	-32.6 (4.4) ^a -26.7 (4.0) -28.5 (4.4) -31.4 (4.3)	-34.1 (4.09)	-37.7 (4.3)
$\Delta\Delta G_{\text{calc}}$ (MM/3D-RISM) (kcal/mol)	0 0 0 0	2.7 ^b -3.5 -3.4 -0.4	-3.7 ^b -9.8 -9.7 -6.7	0 0 0 0	-1.5 ^b -7.4 -5.6 -2.7	-5.1 ^b -11.0 -9.2 -6.3
$\Delta\Delta G_{37}^{\circ}$ (kcal/mol) (Experiment)	0	-0.86		0	-1.76	
Energies corresponding to the molecular mechanics base pairing and base stacking interactions						
Base pairing energy (kcal/mol)	-13.2 (1.8) ^a -5.8 (1.4) -1.8 (0.7) -12.5 (1.7)	-7.9 (1.7)	-9.5 (2.4)	-8.3 (4.7) ^a -5.9 (1.4) -1.5 (1.0) -12.8 (1.6)	-5.2 (1.5)	-9.2 (2.9)
Stacking energy Step 1 (kcal/mol)	-9.9 (1.7) ^a -9.7 (2.2) -6.5 (1.8) -12.6 (3.6)	-5.4 (1.9)	-9.4 (1.69)	-11.3 (4.6) ^a -5.4 (2.2) -5.0 (2.2) -10.6 (2.6)	-4.8 (1.7)	-6.9 (2.1)
Stacking energy Step 2 (kcal/mol)	-14.3 (2.3) ^a -8.8 (2.6) -12.6 (2.1) -13.6 (2.1)	-14.8 (2.4)	-13.1 (1.8)	-9.9 (3.4) ^a -7.9 (2.1) -8.3 (1.8) -8.4 (2.2)	-10.1 (1.7)	-6.7 (1.9)

The values within parentheses correspond to standard deviations. $\Delta\Delta G_{\text{calc}}(7\text{G-A}) = \Delta G(7\text{G-A}) - \Delta G(\text{G-A})$. a - these numbers refer to the different geometries of G-A base pairs in the unmodified duplex. b - $\Delta\Delta G_{\text{calc}}$ calculated relative to the reference unmodified duplexes with different geometries of G-A base pair

(B) Energetics of the duplexes with 7G-G pair (energies in kcal/mol)

Duplexes	GGC (G-G) Anti-Syn Syn-Anti	G7GC (7G(N1H)-G) (GM1)	G7GC (7G(N1H)-G) (GM2)	G7GC (7G(N3H)-G) (GM3)	CGG (G-G) Anti-Syn Syn-Anti	C7GG (7G(N1H)-G) (GM1)	C7GG (7G-G) (GM2))	C7GG (7G(N3H)-G) (GM3)
Hybridization energies (ΔG) and changes in hybridization energies ($\Delta\Delta G$) upon modification								
ΔG (MM/3D-RISM) (kcal/mol)	-30.7 (4.3) ^a -30.5 (4.4)	-35.8 (4.5)	-34.6 (4.5)	-40.9 (4.2)	-31.6 (4.3) ^a -30.2 (4.3)	-35.4 (4.2)	-35.3 (4.2)	-40.2 (4.1)
$\Delta\Delta G_{\text{calc}}$ (MM/3D-RISM) (kcal/mol)	0 0	-5.1 ^b -5.3	-3.9 ^b -4.1	-10.2 ^b -10.4	0 0	-3.9 ^b -5.2	-3.7 ^b -5.1	-8.6 ^b -9.9
$\Delta\Delta G^{\circ}_{37}$ (kcal/mol) (Experiment)	0	-0.95			0	-1.76		
Energies corresponding to the molecular mechanics base pairing and base stacking interactions								
Base pairing energy (kcal/mol)	-10.2 (2.6) ^a -10.2 (2.8)	-21 (2.9)	-12.0 (2.3)	-24.2 (2.8)	-13.8 (2.4) ^a -13.5 (2.6)	-19.6 (3.2)	-12.4 (2.2)	-23.9 (2.9)
Stacking energy Step 1 (kcal/mol)	-4.0 (2.1) ^a -11.6 (2.6)	-11.6 (1.9)	-14.2 (2.8)	-17.1 (1.8)	-13.6 (3.1) ^a -5.6 (2.5)	-12.8 (1.9)	-9.6 (1.9)	-9.9 (1.8)
Stacking energy Step 2 (kcal/mol)	-11.5 (2.5) ^a -3.9 (2.2)	-11.2 (1.8)	-13.2 (2.1)	-8.1 (1.6)	-5.6 (2.4) ^a -13.8 (3.1)	-10.2 (2.2)	-16 (1.8)	-12.2 (2.1)

The values within parentheses correspond to standard deviations. $\Delta\Delta G_{\text{calc}}(7\text{G-G}) = \Delta G(7\text{G-G}) - \Delta G(\text{G-G})$. a - these numbers refer to the different geometries of G-G base pairs in the unmodified duplex. b - $\Delta\Delta G_{\text{calc}}$ calculated relative to the reference unmodified duplexes with different geometries of G-G base pair.

Table S3. (A) Occurrences (in %) of hydrogen bonds between the 7G-A base pair observed in MD simulations

Duplex	Hydrogen bonds (Donor–Hydrogen---Acceptor)	Occurrence (%)
Duplex - G7GC (7G(N1H)-A) (GM1)	(7G5)N1-HN1---N1(A14)	61
Duplex - G7GC (7G(N1H,N3H,N2-imino)-A) (GM3)	(7G5)N1-HN1---N1(A14)	62
	(A14)N6-H62---N2(7G5)	49
Duplex - C7GG (7G(N1H)-A) (GM2)	(7G5)N2-H21---N3(A14)	44
Duplex - C7GG (7G(N1H,N3H,N2-imino)-A) (GM3)	(7G5)N1-HN1---N1(A14)	68
	(A14)N6-H62---N2(7G5)	46

(B) Occurrences (in %) of hydrogen bonds between the 7G-G base pair observed in MD simulations

Duplex	Hydrogen bonds (Donor–Hydrogen---Acceptor)	Occurrence (%)
Duplex - G7GC (7G(N1H)-G) (GM1)	(7G5)N1-HN1---O6(G14)	88

	(G14)N1-H1---O6(7G5)	59
Duplex - G7GC (7G(N1H)-G) (GM2)	(G14)N2-H21---N3(7G5)	73
	(7G5)N2-H22---N3(G14)	12
Duplex - G7GC (7G(N3H)-G) (GM3)	(G14)N2-H21---O6(7G5)	88
	(G14)N1-H1---N1(7G5)	76
	(7G5)N2-H21---O6(G14)	65
Duplex - C7GG (7G(N1H)-G) (GM1)	(7G5)N1-HN1---O6(G14)	90
	(G14)N1-H1---O6(7G5)	46
Duplex - C7GG (7G(N1H)-G) (GM2)	(G14)N2-H21---N3(7G5)	72
	(7G5)N2-H22---N3(G14)	21
Duplex - C7GG (7G(N3H)-G) (GM3)	(G14)N2-H22---O6(7G5)	88
	(G14)N1-H1---N1(7G5)	73
	(7G5)N2-H21---O6(G14)	63

Table S4. ^{13}C NMR chemical shifts for 7G and G (in ppm).

	C6	C2	C4	C8	C5	C1'	C4'	C2'	C3'	C5'
7G	154.36	152.91	160.71	142.44	107.73	89.18	85.24	74.48	69.75	61.20
G ^a	157.82	154.66	152.33	136.67	117.72	87.46	86.26	74.75	71.43	62.45

Table S5. ^1H NMR chemical shifts for 7G and G (in ppm).

	H8	NH ₂	N ¹ H	H1'	H2'	H3'	H4'	H5'/H5''	2'OH/3'OH	5'OH
7G	8.28 (s) ^c	6.19 (s)	10.90 (brs)	5.96 (d)	4.35 (q)	4.06 (q)	3.88 (q)	3.65 (dt) 3.53 (ddd)	5.37 (d) 5.12 (d)	5.04 (t)
G ^b	7.97	6.52	10.75	5.72	4.43	4.11	3.90	3.64 3.55	5.45 5.20	5.10

a: SDBSWeb; https://sdb.db.aist.go.jp/sdb/cgibin/direct_frame_disp.cgi?sdbno=831&spectrum_type=CNMR&fname=CDS03115

b: SDBSWeb https://sdb.db.aist.go.jp/sdb/cgi-bin/direct_frame_disp.cgi?sdbno=831&spectrum_type=HNMR&fname=HSP49445

c: peak multiplicity labels: s - singlet, brs - broad singlet, d - doublet, t - triplet, q - quartet, dt - doublet of triplets, ddd - doublet of doublets of doublets

Table S6. Conformational preferences of 7G-N1H nucleoside

Conformational properties	Sugar Pucker		Base orientation		γ torsion
	SOUTH population (%)	NORTH population (%)	SYN population (%)	ANTI population (%)	g^+ population (%)
FF99_ χ_{KOL0} _ γ_{bsc0}	55.6	44.4	44.54	55.25	93.96
NMR (Baranowski et al. 2022)	62-67 ^a	33-38 ^a	ANTI		g^+
	59 ^b	41 ^b			62 ^c

^a Reported in (Baranowski et al. 2022).

^b Calculated from coupling constants ($J_{1'2'}/J_{3'4'}$) by the formulas %N = 100[$J_{3'4'}/(J_{1'2'} + J_{3'4'})$], %S = 100[$J_{1'2'}/(J_{1'2'} + J_{3'4'})$] from experimental (NMR) data reported in (Baranowski et al. 2022).

^c Calculated from coupling constants $J_{4'5'}=3.80$ Hz $J_{4'5''}=3.70$ Hz by the formulas $gg+gt+tg=1$, $J_{4'5'}=2.4gg+2.6gt+10.6tg$, $J_{4'5''}=1.3gg+10.5gt+3.8tg$

Supporting Figures

Figure S1. The aromatic/amino region of ^1H NMR spectrum of the **7G-A** containing duplexes was recorded at different temperatures with the resonance belonging to the A14 amino group marked.

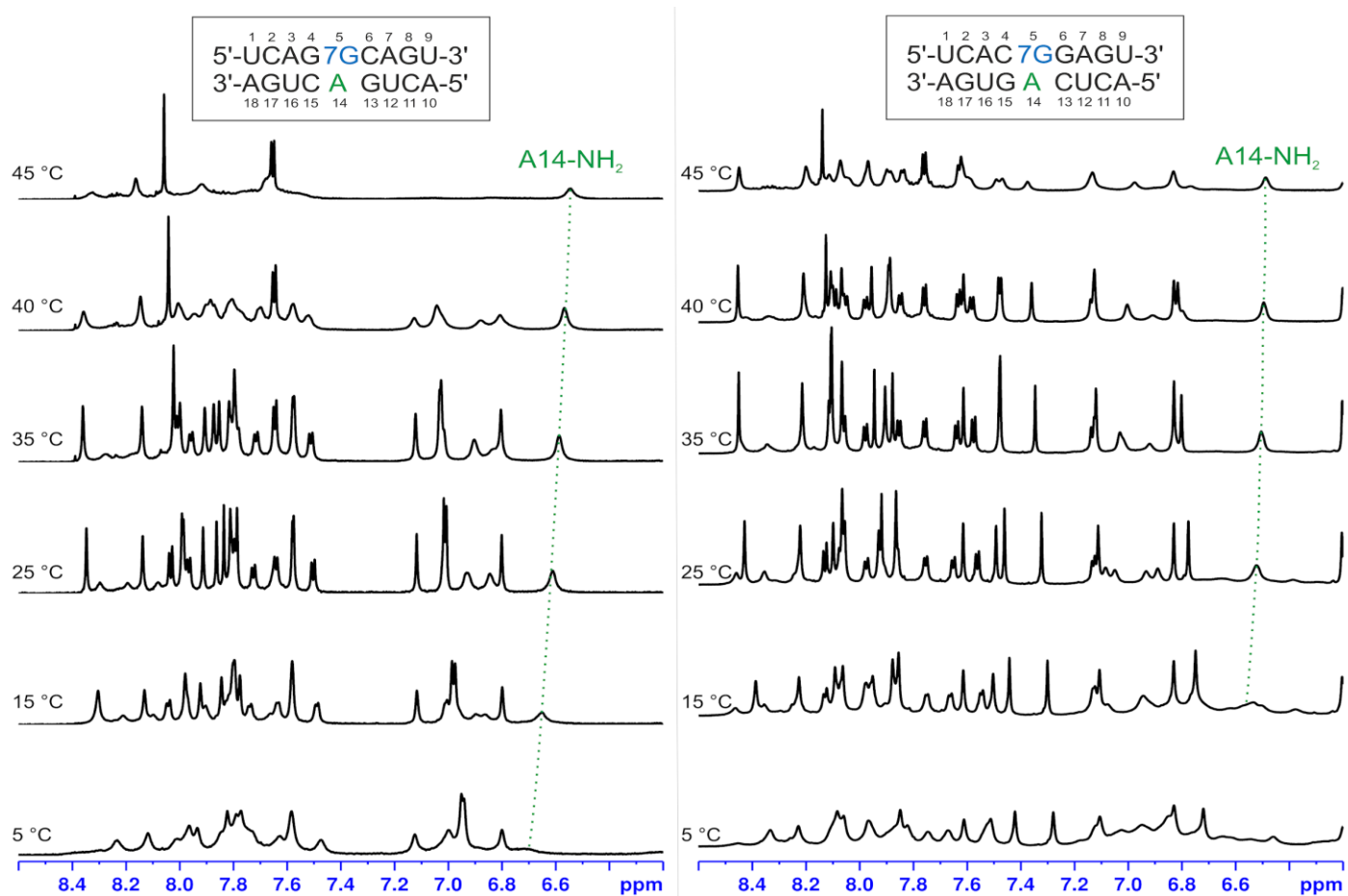


Figure S2. The amino portion of the ^1H - ^{15}N SOFAST-HMQC spectra of the **7G-A** containing duplexes. Positive contour levels in black, negative in red. Some small negative peaks visible are due to a limited signal-to-noise ratio.

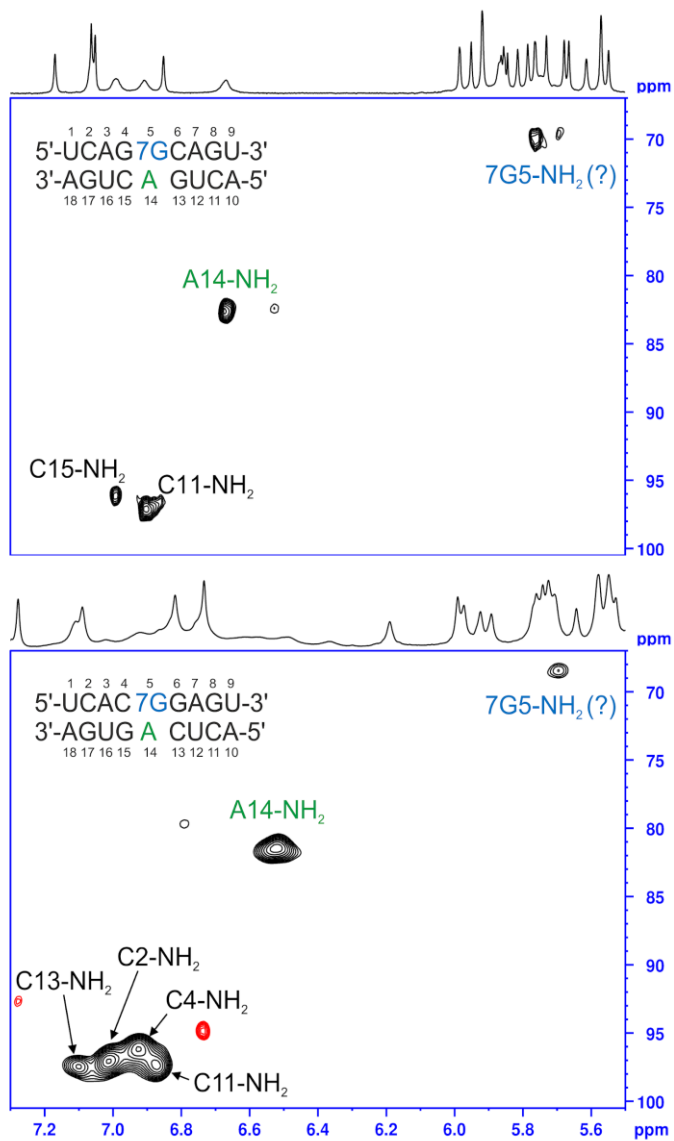


Figure S3. The imino regions of the standard (top) and ^{15}N -selective ^1H NMR spectra of one of the duplexes containing the 7G-G mismatch with the residue G14 site-specifically enriched in ^{15}N . Spectra recorded at 25 °C. Resonances belonging to a second, minor spectral form marked in red stars.

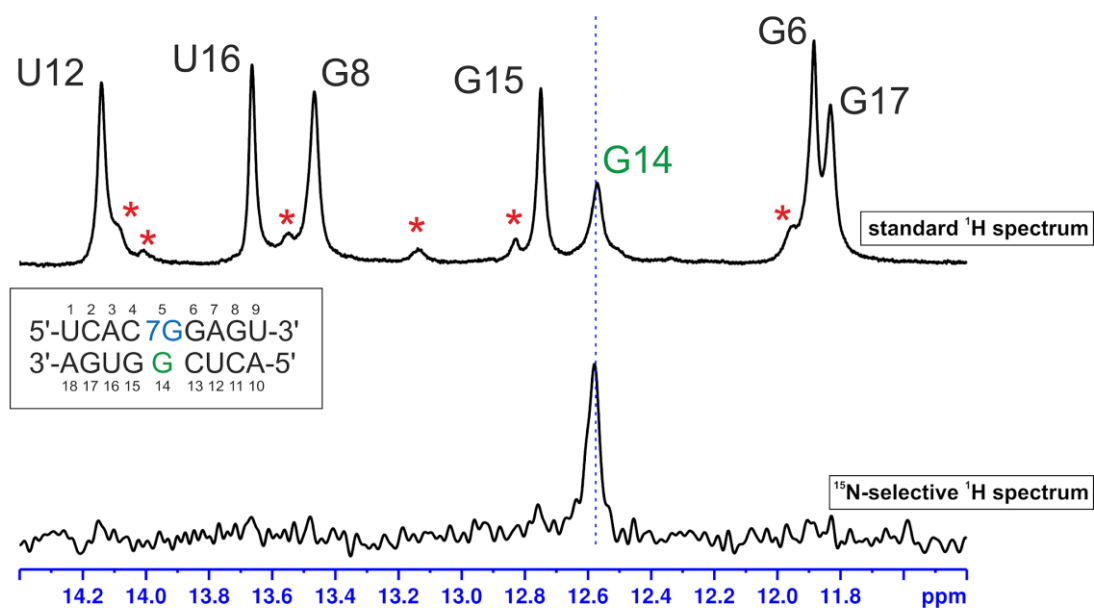


Figure S4. Structures of the tautomers of (A) m⁹Gua, (B) m⁷Gua studied in this work: Keto-N1H (T1), Keto-N3H (T2), Cis-enol (T3), Trans-enol (T4), N1H,N3H,N2-imino-1 (T5) and N1H,N3H,N2-imino 2 (T6).

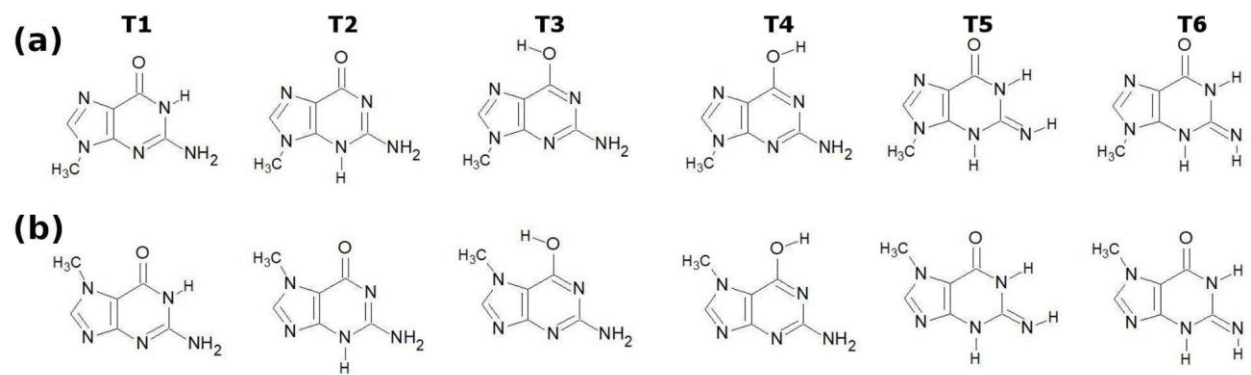


Figure S5. Comparison of the Zero-point-energy (ZPE)-corrected relative free energies of m⁷Gua, m⁹Gua tautomers with respect to keto-N1H(T1) (the most stable tautomer) in the (a) gas phase and (b) in the solvent phase.

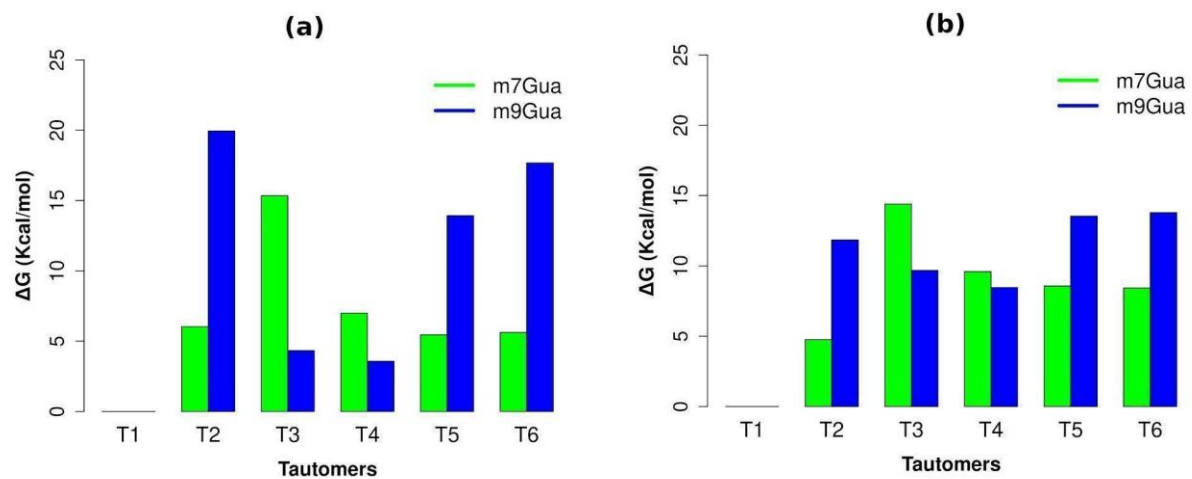
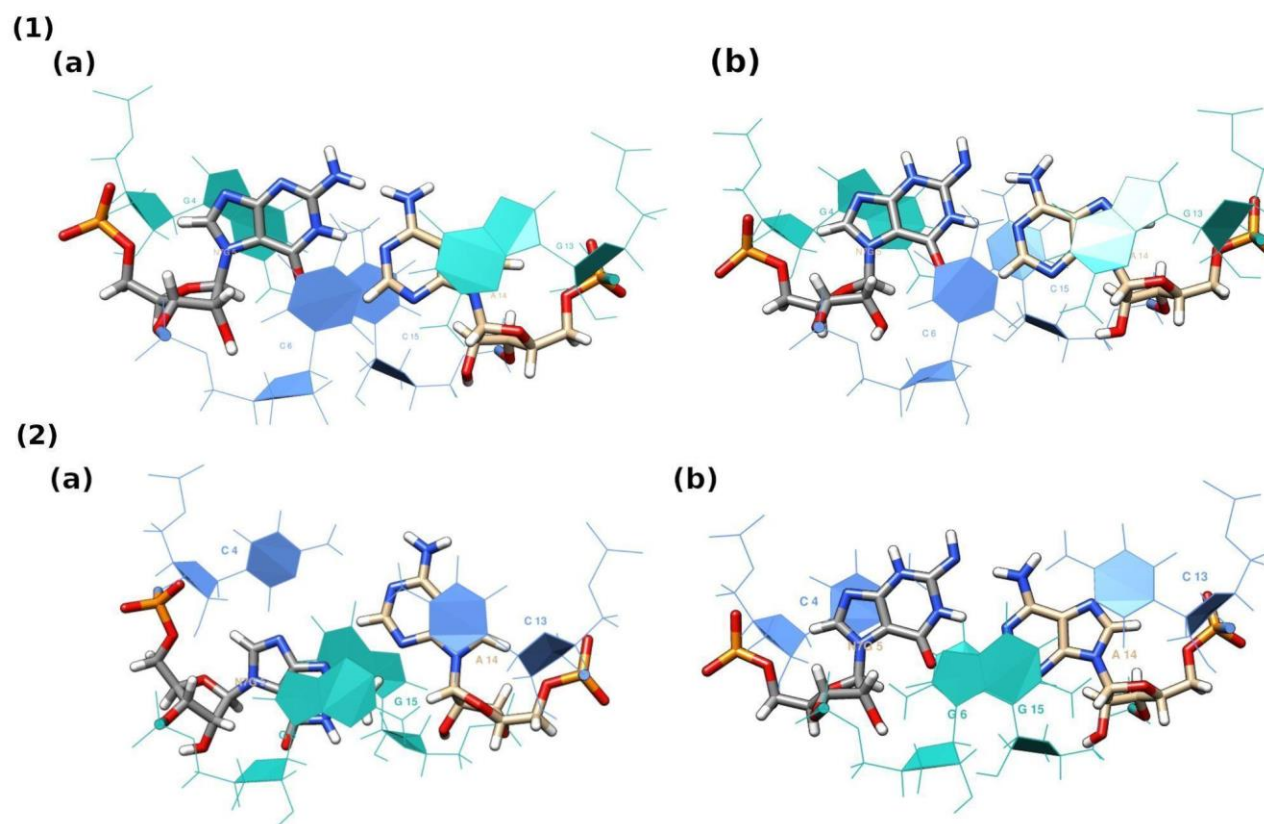


Figure S6. (A) Observed stacked geometries of (1) (a) 5'G7G(N1H)C-3'CAG (GM1) and (b) 5'G7G(N1H,N3H,N2-imino)C-3'CAG (GM3) and (2) (a) 5'C7G(N1H)G-3'GAC (GM2) and (b) 5'C7G(N1H,N3H,N2-imino)G-3'GAC (GM3). 7G5 and A14 residues are shown in sticks (carbons are shown in grey for 7G15 and in tan for A14, nitrogens in blue, oxygens in red and hydrogens in white), for residues 4,6,13 and 15, G-residues are shown in light seagreen, and C-residues are shown in cornflower blue color with base pair 4-15 in the back, 5-14 in the middle and 6-13 in the front.



(B) Observed stacked geometries of (a) 7G(N1H)-G (GM1); (b) 7G(N1H)-G (GM2); (c) 7G(N3H)-G (GM3) for (1) 5'G7GC-3'CGG and (2) 5'C7GG-3'GGC contexts. 7G5 and G14 residues are shown in sticks (carbons are shown in grey for 7G15 and in tan for A14, nitrogens in blue, oxygens in red and hydrogens in white), for residues 4,6,13 and 15, the G-residues are shown in light seagreen, and C-residues are shown in cornflower blue color with base pair 4-15 in the back, 5-14 in the middle and 6-13 in the front.

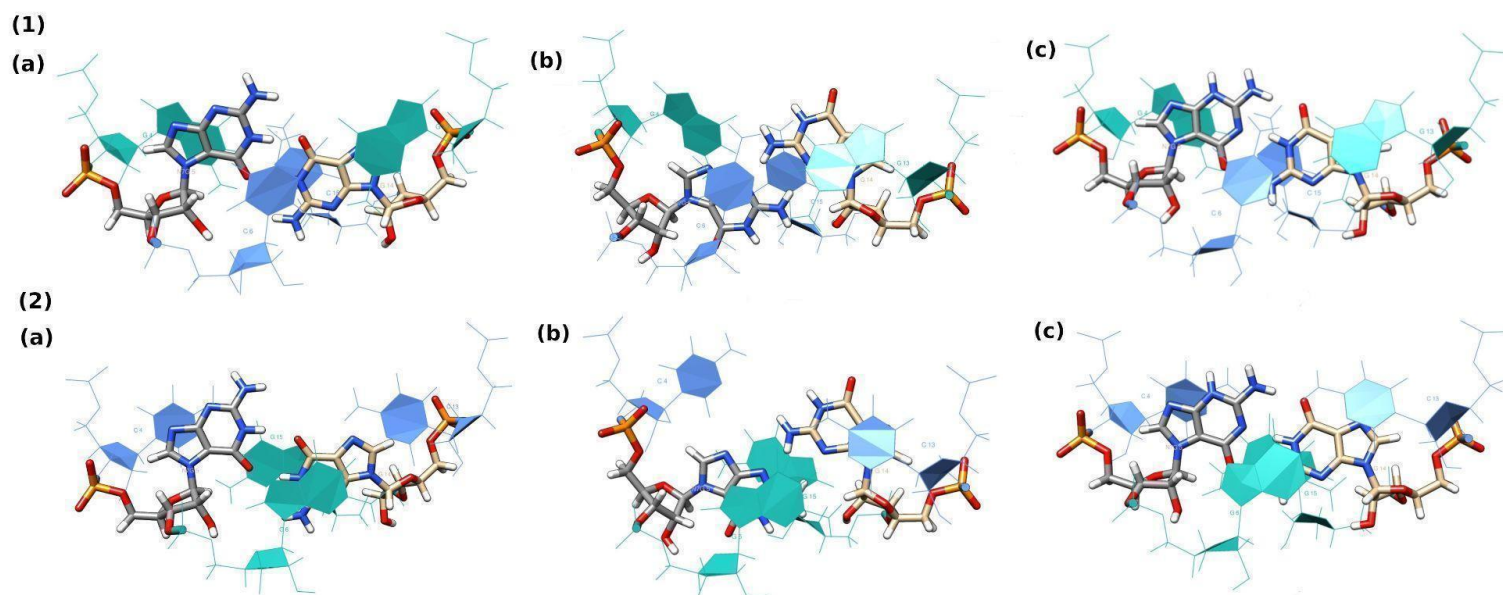
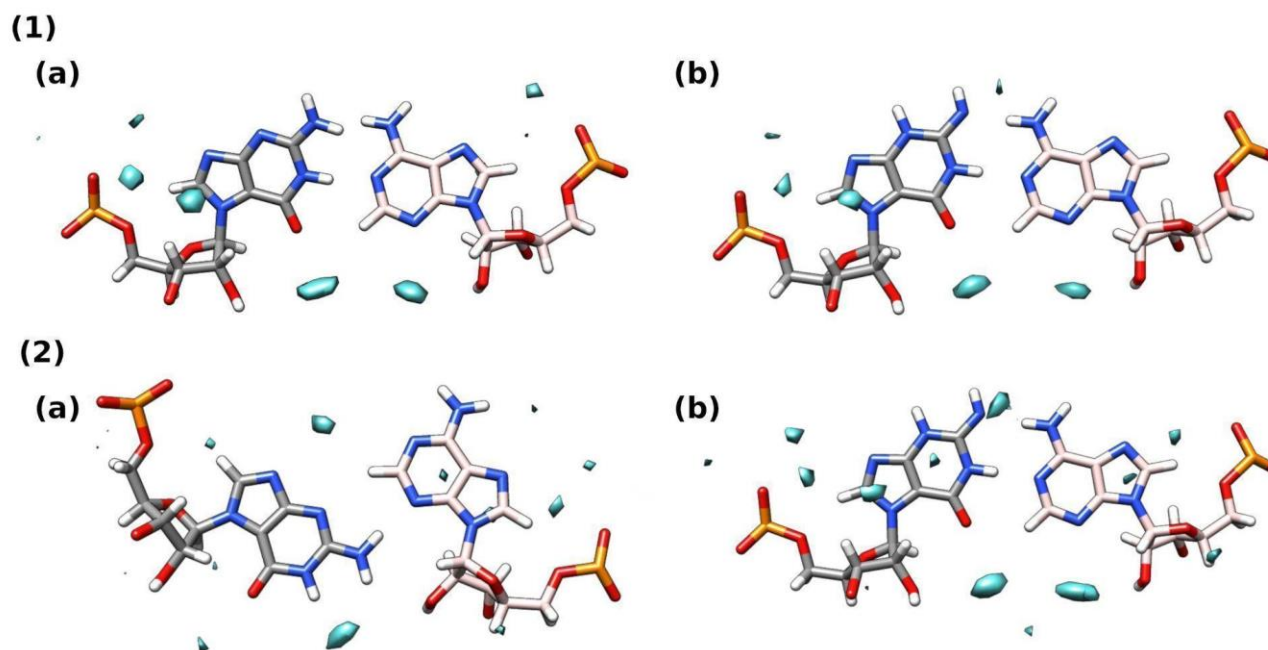


Figure S7. (A) Observed water occupancy maps around (1) (a) 5'G7G(N1H)C-3'CAG (GM1) and (b) 5'G7G(N1H,N3H,N2-imino)C-3'CAG (GM3) and (2) (a) 5'C7G(N1H)G-3'GAC (GM2) and (b) 5'C7G(N1H,N3H,N2-imino)G-3'GAC (GM3). 7G is shown in grey.



(B) Observed water occupancy maps around (a) 7G(N1H)-G (GM1); (b) 7G(N1H)-G (GM2); (c) 7G(N3H)-G (GM3) for (1) 5'G7GC-3'CGG and (2) 5'C7GG-3'GGC contexts. 7G is shown in grey.

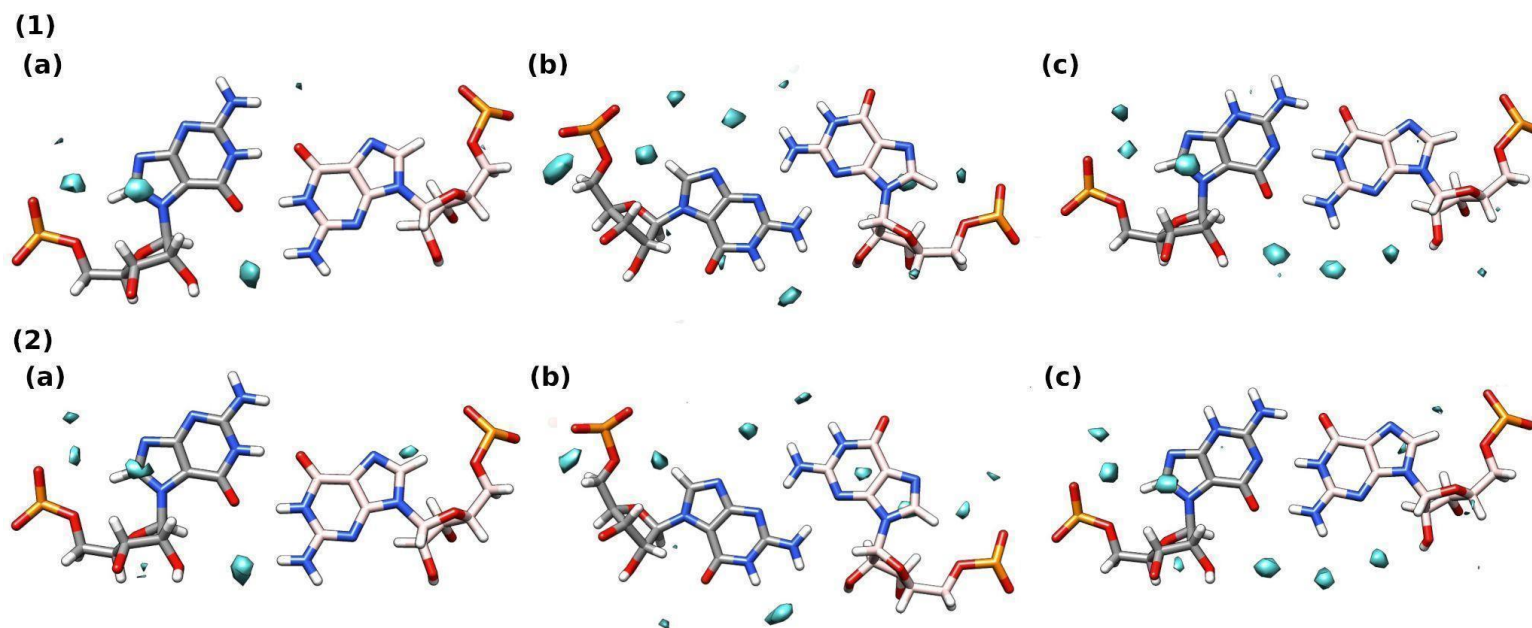


Figure S8. ^1H spectrum of 7G in DMSO- d_6 at 25 $^\circ\text{C}$

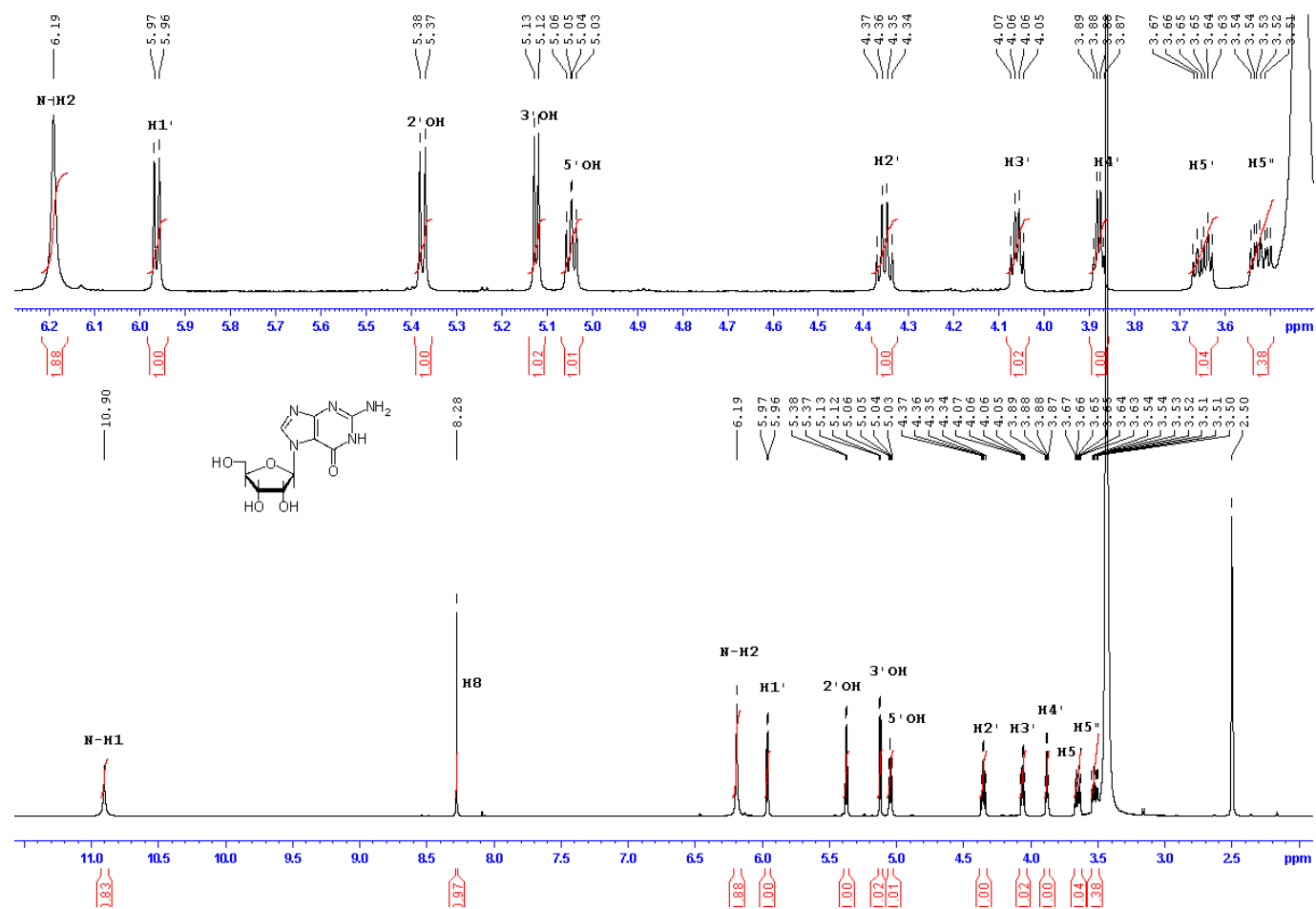


Figure S9. ^{13}C spectrum of 7G in DMSO- d_6 at 25 $^{\circ}\text{C}$

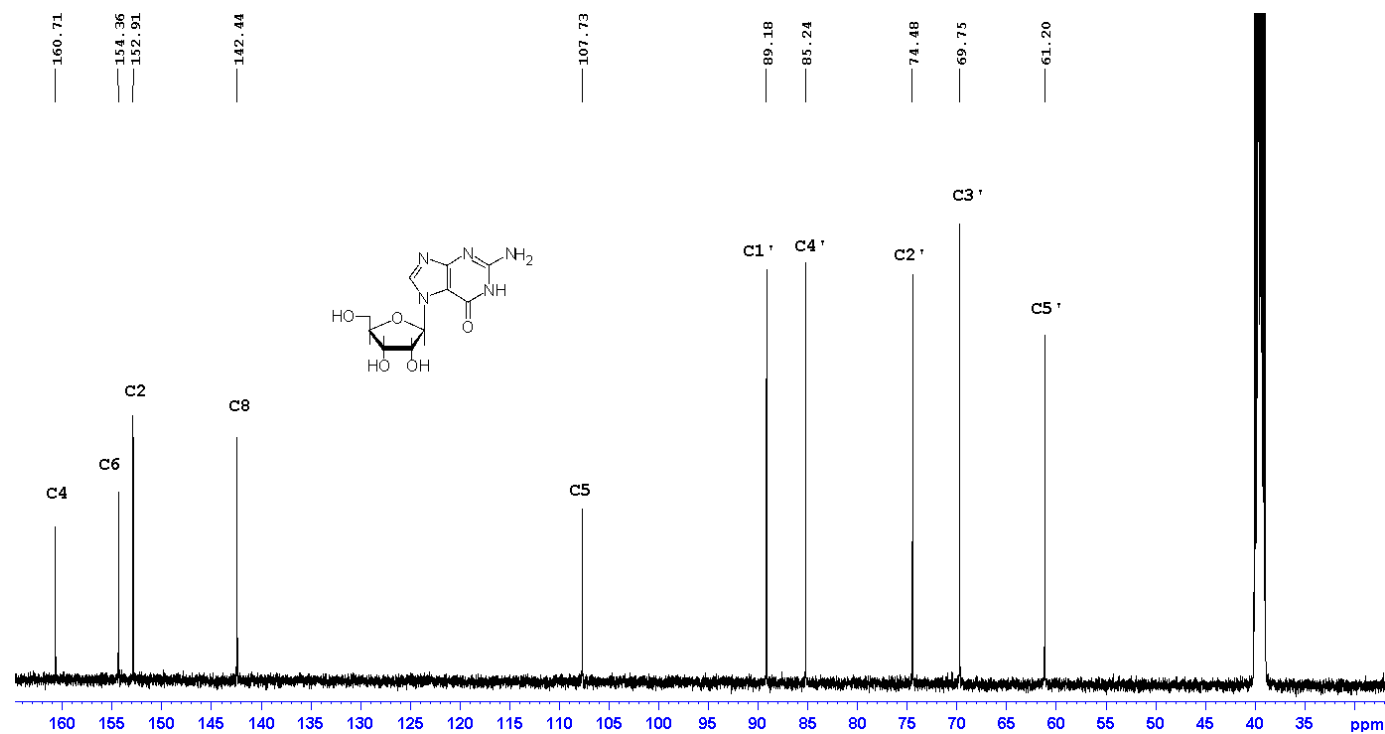


Figure S10. ^1H - ^1H COSY spectrum of 7G in DMSO- d_6 at 25 °C

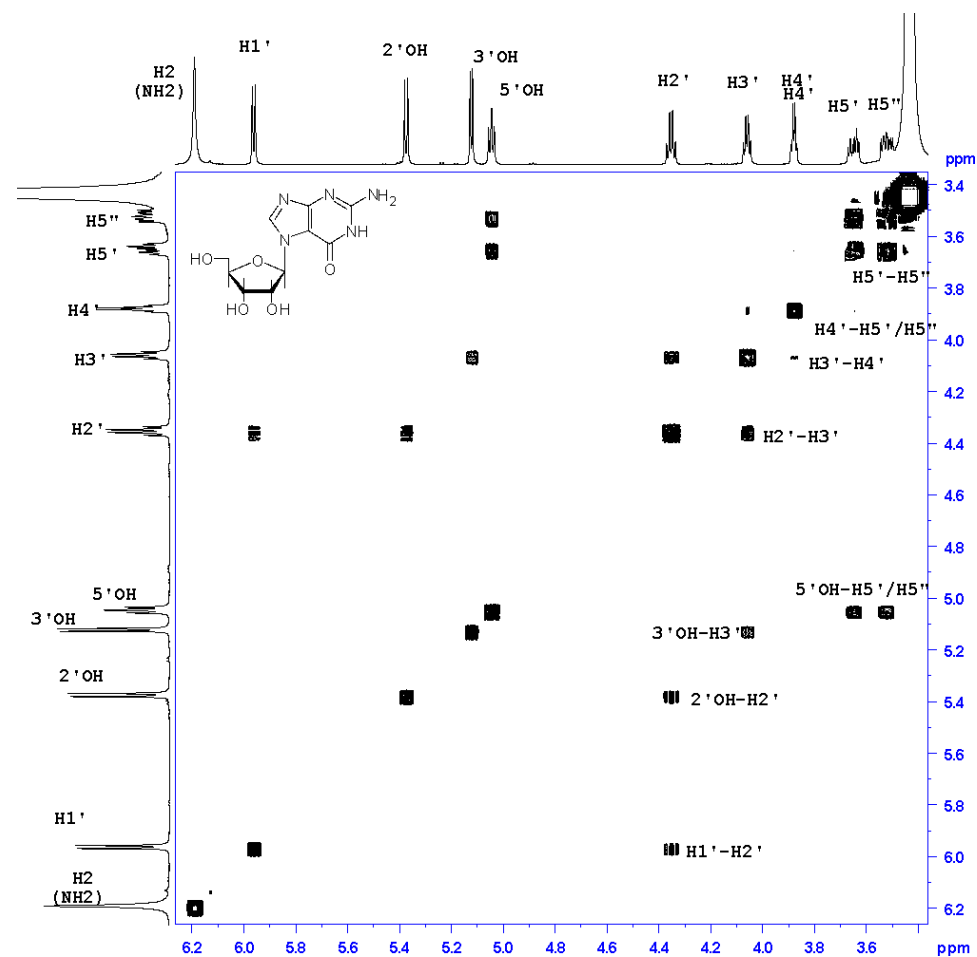


Figure S11. ^1H - ^{13}C gHSQC spectrum of 7G in DMSO- d_6 at 25 °C

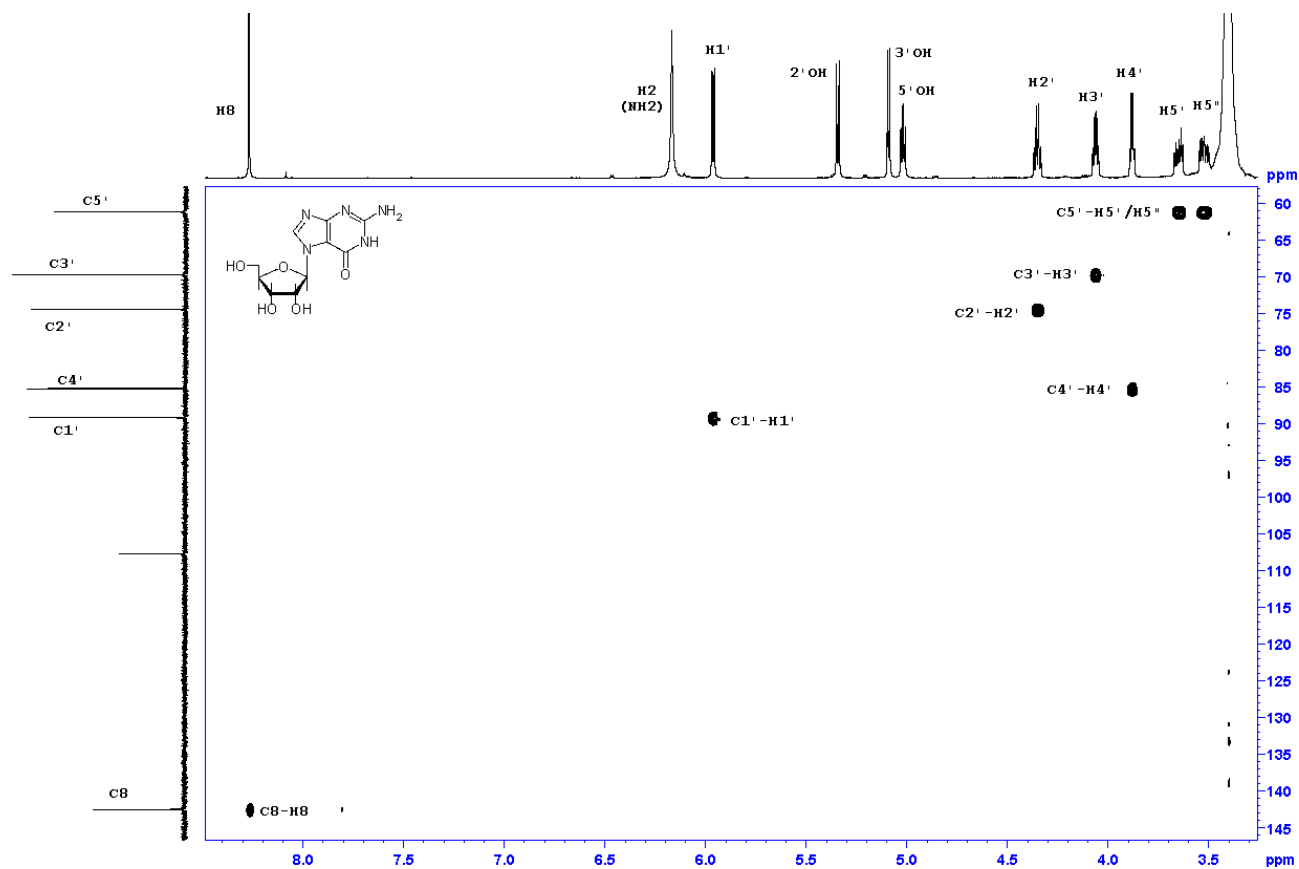


Figure S12. ^1H - ^{13}C gHMBC spectrum of 7G in DMSO- d_6 at 25 °C

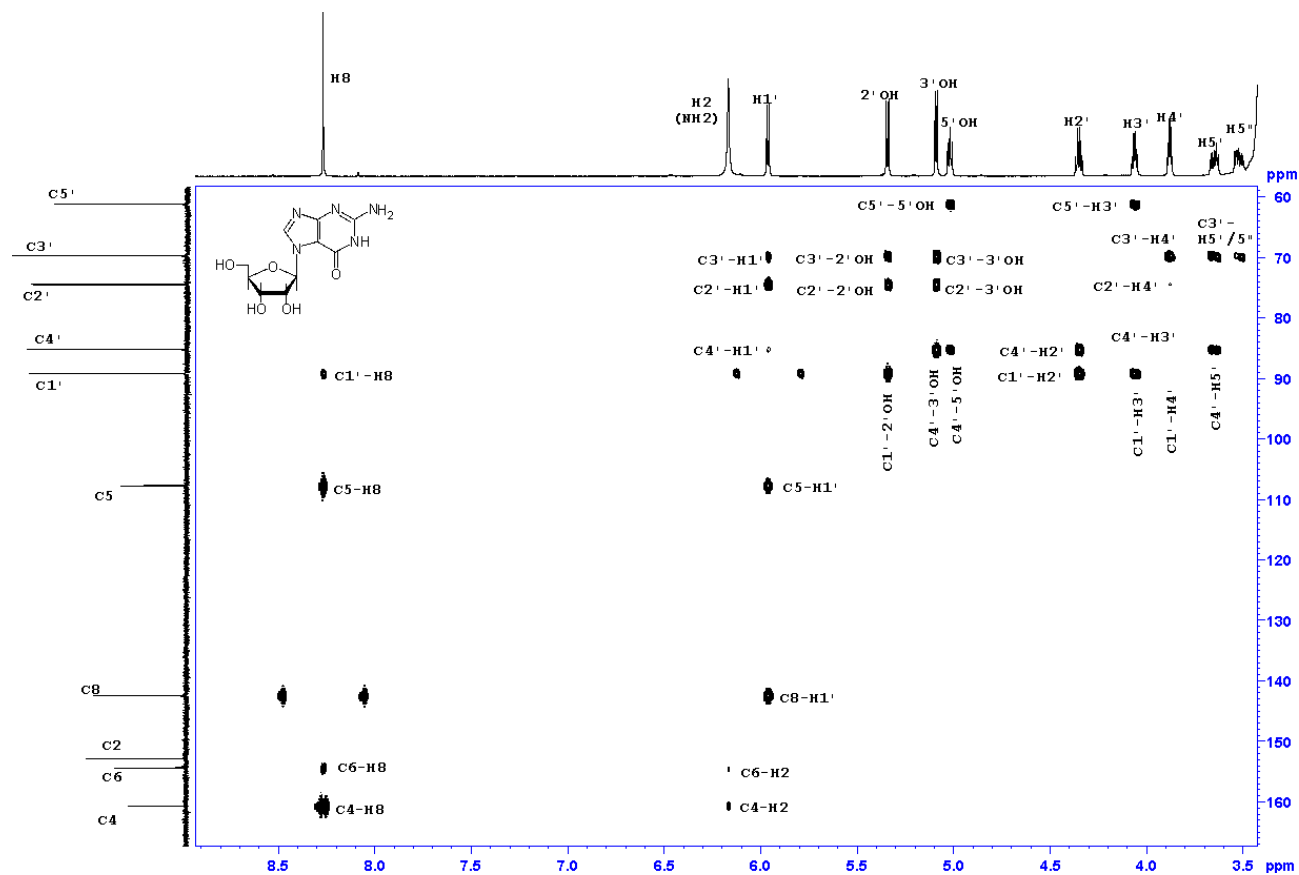


Figure S13. ^1H - ^{15}N gHSQC spectrum of 7G in DMSO- d_6 at 25 °C

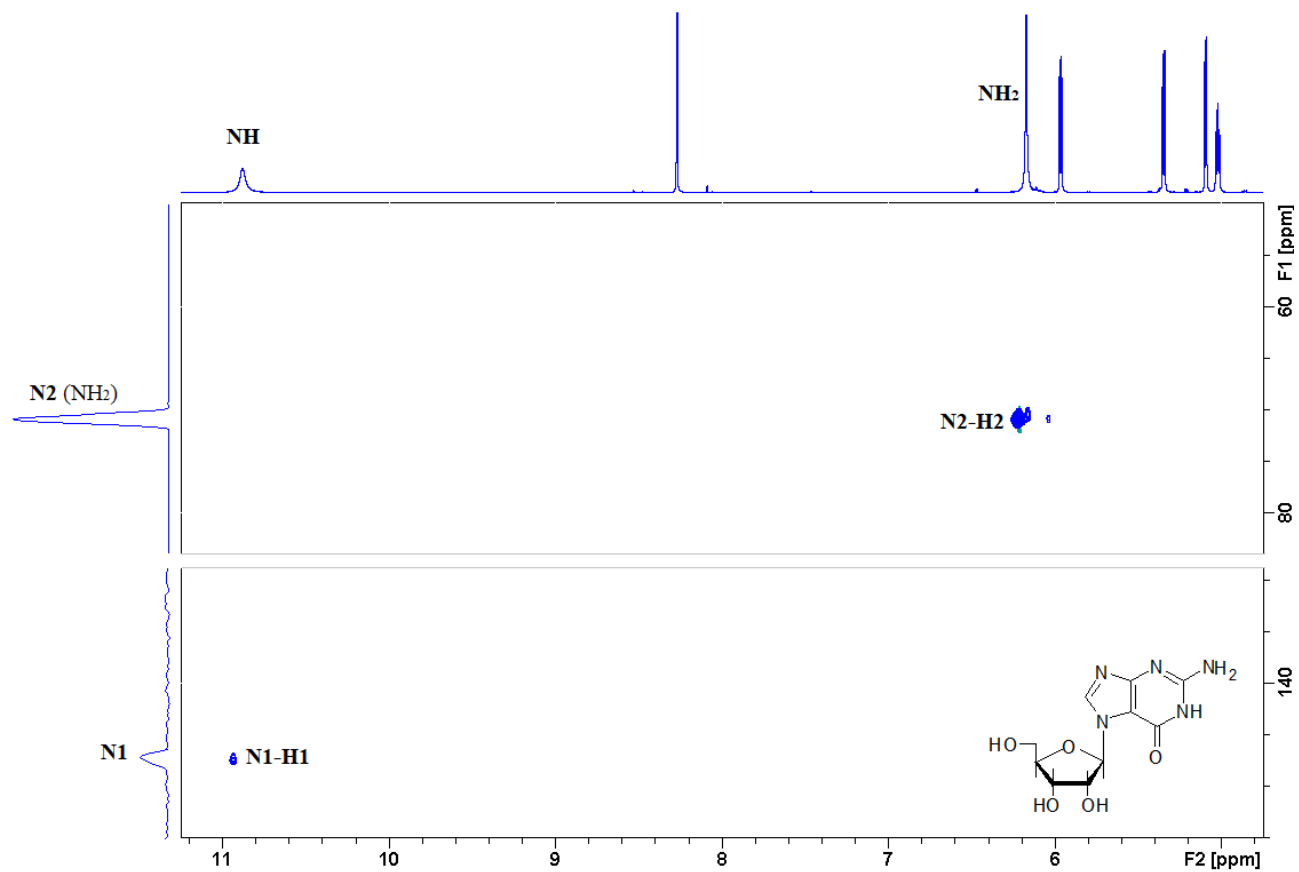


Figure S14. ^1H - ^{15}N gHMBC spectrum of 7G in DMSO- d_6 at 25 °C

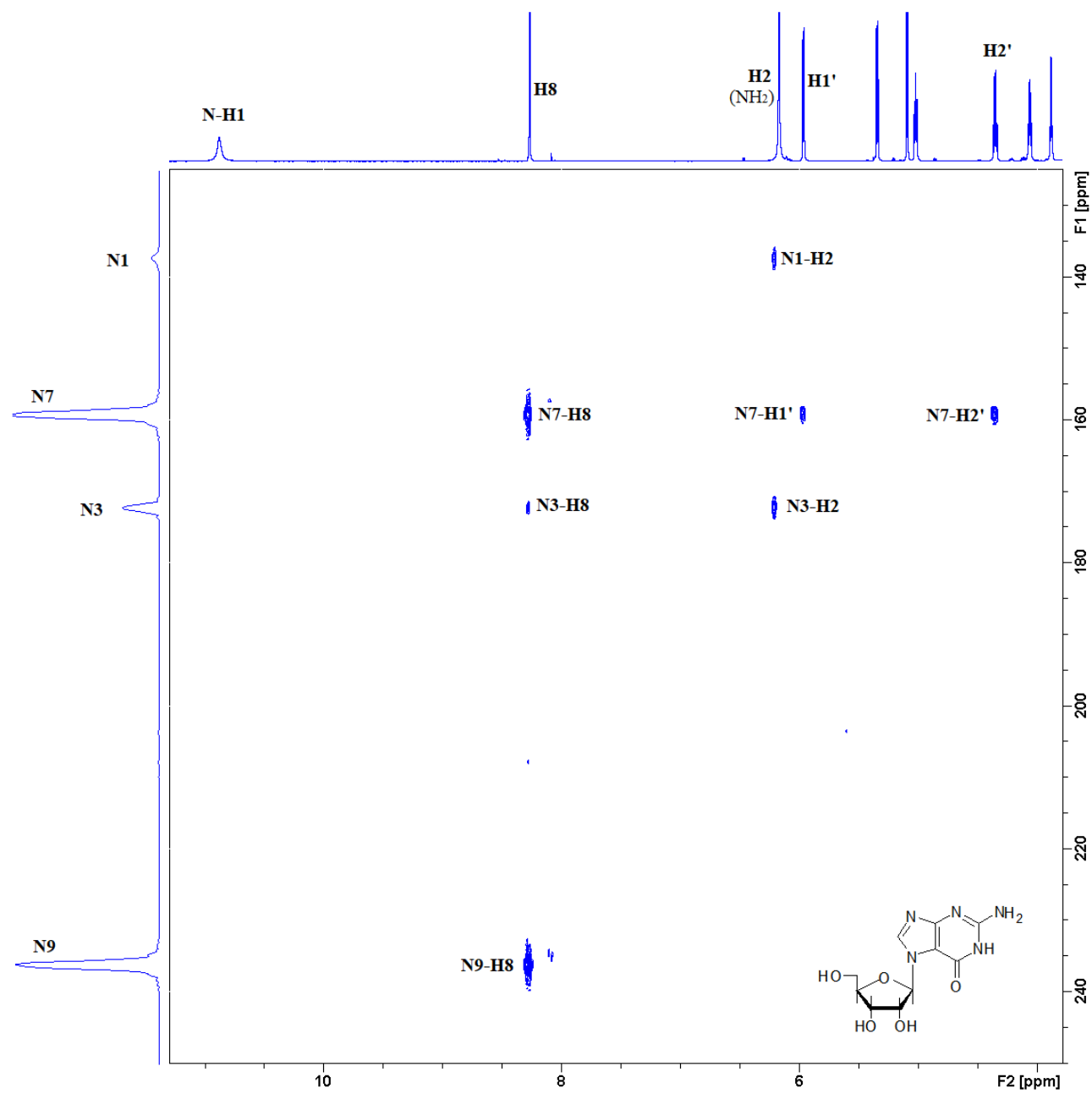


Figure S15. Quantum mechanical energy profiles around χ torsional angles (O4'-C1'-N7-C5) for 7G-N1H corresponding to the four conformational schemes, respectively.

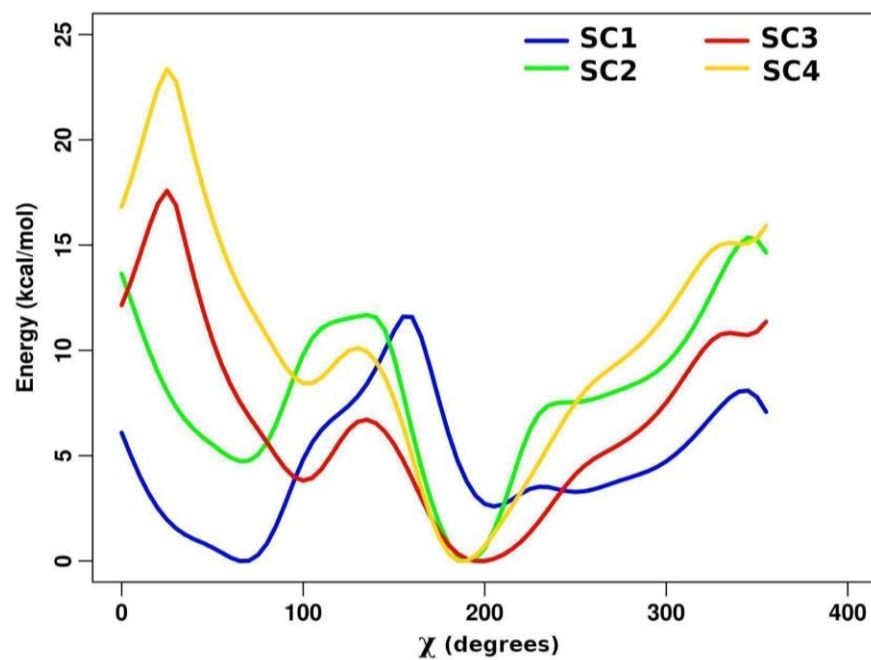


Figure S16. Energy profiles of the $\chi+180^\circ$ (O4'-C1'-N7-C8) torsional angles for 7G corresponding to QM calculations (black), MM calculations with the FF99 parameter sets keeping the glycosidic torsion parameters zero (red) and MM calculations with the FF99 parameter sets combined with the newly derived χ torsional parameters and the newly developed partial atomic charges (FF99_ χ_{KOL0}) (green) by fitting the difference between the QM and MM energies for (a) SC1, (b) SC2, (c) SC3 and (d) SC4 respectively. The minimum energies were set to zero for convenience.

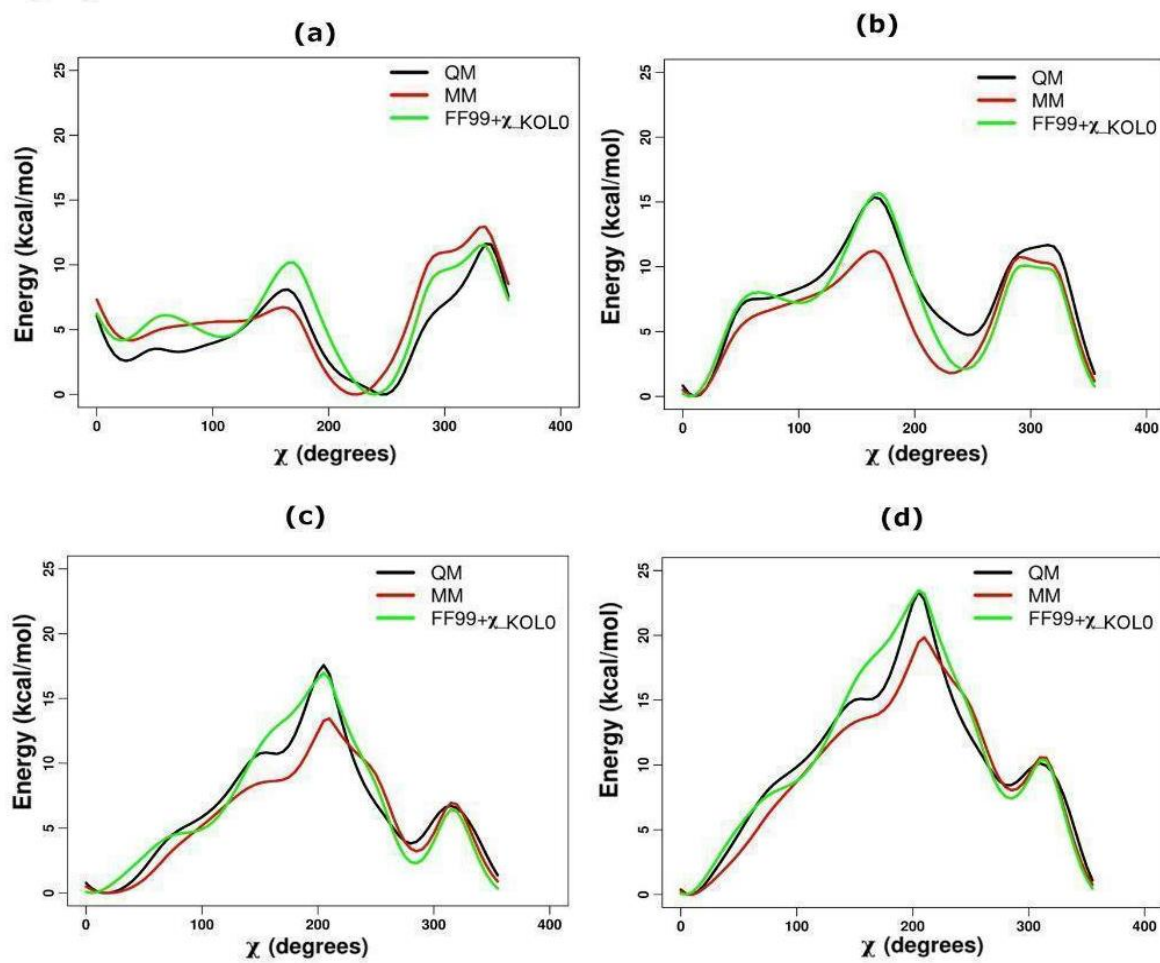
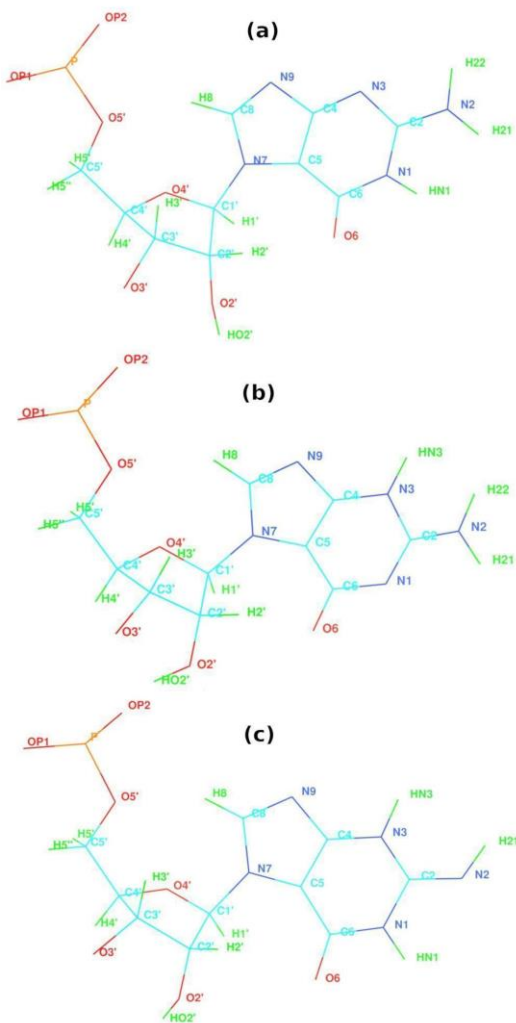


Figure S17. Representative diagrams of the 7G tautomers (a) 7G-N1H; (b) 7G-N3H and (c) 7G-N1H,N3H,N2-imino in their nucleotide forms with the atom names.



APPENDIX 1

AMBER preparatory file for 7G-N1H

0 0 2

Developed at Institute of Bioorganic Chemistry PAS by Nivedita Dutta

7G.res

7G INT 0

CORRECT OMIT DU BEG

0.0000

1	DUMM	DU	M	0	-1	-2	0.000	.0	.0	.00000
2	DUMM	DU	M	1	0	-1	1.449	.0	.0	.00000
3	DUMM	DU	M	2	1	0	1.523	111.21	.0	.00000
4	P	P	M	3	2	1	1.609	119.956	-133.992	1.1662
5	OP1	O	E	4	3	2	1.485	107.081	145.987	-0.7760
6	OP2	O	E	4	3	2	1.486	109.428	15.165	-0.7760
7	O5'	OS	M	4	3	2	1.593	103.856	-99.934	-0.4989
8	C5'	CI	M	7	4	3	1.426	120.975	-62.582	0.0558
9	H5'	H1	E	8	7	4	1.091	109.725	30.107	0.0679
10	H5"	H1	E	8	7	4	1.090	109.765	-89.566	0.0679
11	C4'	CT	M	8	7	4	1.511	110.226	150.387	0.1065
12	H4'	H1	E	11	8	7	1.091	107.708	175.369	0.1174
13	O4'	OS	M	11	8	7	1.454	109.053	-60.938	-0.3548
14	C1'	CT	M	13	11	8	1.418	109.791	140.832	0.1358
15	N7	N*	S	14	13	11	1.463	108.323	-115.257	-0.1054
16	C5	CB	S	15	14	13	1.374	126.614	-139.229	-0.1525
17	C6	C	B	16	15	14	1.344	127.448	-0.092	0.4915
18	N1	NA	B	17	16	15	1.332	110.623	-179.820	-0.5096
19	C2	CA	B	18	17	16	1.338	129.242	-0.105	0.6987
20	N3	NC	S	19	18	17	1.350	118.594	0.063	-0.6296
21	C4	CB	S	20	19	18	1.405	117.753	0.077	0.5754
22	N9	NB	S	21	20	19	1.388	132.274	179.802	-0.6217
23	C8	CK	S	22	21	20	1.311	103.867	-179.994	0.2691
24	H8	H5	E	23	22	21	1.080	123.107	179.801	0.1270
25	N2	N2	B	19	18	17	1.332	120.712	-179.953	-0.8829
26	H21	H	E	25	18	17	1.010	119.949	0.027	0.3901
27	H22	H	E	25	18	17	1.009	119.979	179.985	0.3901

28	HN1	H	E	18	17	16	1.091	115.414	179.954	0.3546
29	O6	O	E	17	16	15	1.239	124.675	0.199	-0.5356
30	H1'	H2	E	14	13	11	1.090	107.041	128.337	0.1210
31	C2'	CT	M	14	13	11	1.529	107.230	7.740	0.0670
32	O2'	OH	S	31	14	13	1.415	108.728	88.062	-0.6139
33	HO2'	HO	E	32	31	14	0.957	106.681	-130.227	0.4186
34	H2'	H1	E	31	14	13	1.092	111.690	-146.692	0.0972
35	C3'	CT	M	31	14	13	1.523	100.960	-28.912	0.2022
36	H3'	H1	E	35	31	14	1.089	112.602	-81.930	0.0615
37	O3'	OS	M	35	31	14	1.425	112.670	157.951	-0.5246

LOOP

C4 C5

N7 C8

C4' C3'

IMPROPER

C6 C4 C5 N7

C5 N1 C6 O6

C6 C2 N1 HN1

N1 N3 C2 N2

C2 H21 N2 H22

C5 N3 C4 N9

H8 N7 C8 N9

C1' C8 N7 C5

DONE

STOP

AMBER preparatory files for 7G-N3H

0 0 2

Developed at Institute of Bioorganic Chemistry PAS by Nivedita Dutta

7G.res

7G INT 0

CORRECT OMIT DU BEG

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2	DUMM	DU	M	1	0	-1	1.449	.0	.0	.00000
3	DUMM	DU	M	2	1	0	1.523	111.21	.0	.00000
4	P	P	M	3	2	1	1.609	119.956	-133.992	1.1662
5	OP1	O2	E	4	3	2	1.485	107.081	145.987	-0.7760
6	OP2	O2	E	4	3	2	1.486	109.428	15.165	-0.7760
7	O5'	OS	M	4	3	2	1.593	103.856	-99.934	-0.4989
8	C5'	CI	M	7	4	3	1.426	120.975	-62.582	0.0558
9	H5'	H1	E	8	7	4	1.091	109.725	30.107	0.0679
10	H5"	H1	E	8	7	4	1.090	109.765	-89.566	0.0679
11	C4'	CT	M	8	7	4	1.511	110.226	150.387	0.1065
12	H4'	H1	E	11	8	7	1.091	107.708	175.369	0.1174
13	O4'	OS	M	11	8	7	1.454	109.053	-60.938	-0.3548
14	C1'	CT	M	13	11	8	1.418	109.791	140.832	0.0896
15	N7	N*	S	14	13	11	1.463	108.323	-115.257	-0.0442
16	C5	CB	S	15	14	13	1.374	126.614	-139.229	-0.1063
17	C6	Q2	B	16	15	14	1.344	127.448	-0.092	0.6844
18	N1	NC	S	17	16	15	1.332	110.623	-179.820	-0.7313
19	C2	CA	B	18	17	16	1.338	129.242	-0.105	0.7943
20	N3	NA	B	19	18	17	1.350	118.594	0.063	-0.5465
21	C4	CB	S	20	19	18	1.405	117.753	0.077	0.3786
22	N9	NB	S	21	20	19	1.388	132.274	179.802	-0.5942
23	C8	Q1	S	22	21	20	1.311	103.867	-179.994	0.2237
24	H8	H5	E	23	22	21	1.080	123.107	179.801	0.1430
25	HN3	HQ	E	20	19	18	1.010	121.123	-179.896	0.3903
26	N2	N2	B	20	19	18	1.332	120.712	-179.953	-0.9297
27	H21	H	E	26	20	19	1.010	119.949	0.027	0.4024
28	H22	H	E	26	20	19	1.009	119.979	179.985	0.4024
29	O6	O	E	17	16	15	1.239	124.675	0.199	-0.5527
30	H1'	H2	E	14	13	11	1.090	107.041	128.337	0.1122
31	C2'	CT	M	14	13	11	1.529	107.230	7.740	0.0670
32	O2'	OH	S	31	14	13	1.415	108.728	88.062	-0.6139
33	HO2'	HO	E	32	31	14	0.957	106.681	-130.227	0.4186
34	H2'	H1	E	31	14	13	1.092	111.690	-146.692	0.0972
35	C3'	CT	M	31	14	13	1.523	100.960	-28.912	0.2022
36	H3'	H1	E	35	31	14	1.089	112.602	-81.930	0.0615
37	O3'	OS	M	35	31	14	1.425	112.670	157.951	-0.5246

AMBER preparatory files for 7G-N1H,N3H,N2-imino

0 0 2

This is a remark line

molecule.res

7G INT 0

CORRECT OMIT DU BEG

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2	DUMM	DU	M	1	0	-1	1.449	.0	.0	.00000
3	DUMM	DU	M	2	1	0	1.523	111.21	.0	.00000
4	P	P	M	3	2	1	1.609	119.956	-133.992	1.1662
5	OP1	O2	E	4	3	2	1.485	107.081	145.987	-0.7760
6	OP2	O2	E	4	3	2	1.486	109.428	15.165	-0.7760
7	O5'	OS	M	4	3	2	1.593	103.856	-99.934	-0.4989
8	C5'	CI	M	7	4	3	1.426	120.975	-62.582	0.0558
9	H5'	H1	E	8	7	4	1.091	109.725	30.107	0.0679
10	H5"	H1	E	8	7	4	1.090	109.765	-89.566	0.0679
11	C4'	CT	M	8	7	4	1.511	110.226	150.387	0.1065
12	H4'	H1	E	11	8	7	1.091	107.708	175.369	0.1174
13	O4'	OS	M	11	8	7	1.454	109.053	-60.938	-0.3548
14	C1'	CT	M	13	11	8	1.418	109.791	140.832	0.1638
15	N7	N*	S	14	13	11	1.463	108.323	-115.257	-0.1049
16	C5	CB	S	15	14	13	1.374	126.614	-139.229	0.0143
17	C6	Q2	B	16	15	14	1.339	126.854	-0.089	0.3474
18	N1	NA	B	17	16	15	1.362	110.006	-179.821	-0.2614
19	C2	CA	B	18	17	16	1.319	129.535	-0.114	0.4061
20	N3	NP	B	19	18	17	1.390	118.313	0.082	-0.2727
21	C4	CB	S	20	19	18	1.395	117.046	0.057	0.2187
22	N9	NB	S	21	20	19	1.388	131.556	179.812	-0.5708
23	C8	Q1	S	22	21	20	1.311	103.867	-179.992	0.2667
24	H8	H5	E	23	22	21	1.080	123.107	179.801	0.1256
25	HN3	HQ	E	20	19	18	1.000	120.135	-179.895	0.3181
26	N2	ND	S	19	18	17	1.251	121.753	-179.948	-0.8397
27	H21	H	E	25	19	18	1.009	120.750	179.991	0.3788
28	HN1	H	E	18	17	16	1.004	114.611	179.895	0.3077
29	O6	O	E	17	16	15	1.229	125.845	0.219	-0.4958
30	H1'	H2	E	14	13	11	1.090	107.041	128.337	0.1141

31	C2'	CT	M	14	13	11	1.529	107.230	7.740	0.0670
32	O2'	OH	S	31	14	13	1.415	108.728	88.062	-0.6139
33	HO2'	HO	E	32	31	14	0.957	106.681	-130.227	0.4186
34	H2'	H1	E	31	14	13	1.092	111.690	-146.692	0.0972
35	C3'	CT	M	31	14	13	1.523	100.960	-28.912	0.2022
36	H3'	H1	E	35	31	14	1.089	112.602	-81.930	0.0615
37	O3'	OS	M	35	31	14	1.425	112.670	157.951	-0.5246

LOOP

C4 C5

N7 C8

C4' C3'

IMPROPER

C6 C4 C5 N7

C5 N1 C6 O6

C2 C4 N3 HN3

N1 N3 C2 N2

C5 N3 C4 N9

H8 N7 C8 N9

C1' C8 N7 C5

DONE

STOP

AMBER parameter files for 7G-N1H

Developed at Institute of Bioorganic Chemistry PAS by Nivedita Dutta

MASS

Q1 12.01 0.360

BOND

Q1-H5 367.0 1.080

Q1-N* 440.0 1.371

Q1-NB 529.0 1.304

ANGLE

H5-Q1-N* 50.0 123.05

H5-Q1-NB	50.0	123.05	
N*-Q1-NB	70.0	113.90	
CB-N*-Q1	70.0	105.40	
CT-N*-Q1	70.0	128.80	
CB-NB-Q1	70.0	103.80	
CT-OS-HO	55.0	108.50	manually added from parm99.dat
CI-OS-HO	55.0	108.50	manually modified from parm99.dat & frcmod.parmbsc0
N*-CB-C	70.0000	134.500	
NB-CB-NC	70.0000	122.000	

DIHE

X-Q1-N*-X	4	6.80	180.0	2.
X-Q1-NB-X	2	20.00	180.0	2.
OS-CT-N*-Q1	1	0.0649123	180.0	-4. chi KOL0
OS-CT-N*-Q1	1	0.434356	180.0	-3. chi KOL0
OS-CT-N*-Q1	1	0.952733	0.0	-2. chi KOL0
OS-CT-N*-Q1	1	1.12182	180.0	1. chi KOL0
CT-CT-N*-Q1	1	0.0797247	180.0	-4. chi KOL0
CT-CT-N*-Q1	1	0.683783	180.0	-3. chi KOL0
CT-CT-N*-Q1	1	0.686632	180.0	-2. chi KOL0
CT-CT-N*-Q1	1	0.808183	0.0	1. chi KOL0

IMPROPER

X-X-Q1-H5	1.1	180.	2.
CB-CT-N*-Q1	1.0	180.	2.

NONBON

Q1	1.9080	0.0860
----	--------	--------

AMBER parameter files for 7G-N3H

Developed at Institute of Bioorganic Chemistry PAS by Nivedita Dutta
MASS

Q1 12.01	0.360
Q2 12.01	0.616
HQ 1.008	0.161

BOND

Q1-H5	367.0	1.080
-------	-------	-------

Q1-N* 440.0 1.371
 Q1-NB 529.0 1.304
 NA-CB 461.0 1.354
 NA-HQ 434.10 1.010
 Q2-O 570.0 1.229
 Q2-NC 457.0 1.358
 Q2-CB 447.0 1.419

ANGLE

H5-Q1-N* 50.0 123.05
 H5-Q1-NB 50.0 123.05
 N*-Q1-NB 70.0 113.90
 CB-N*-Q1 70.0 105.40
 CT-N*-Q1 70.0 128.80
 CB-NB-Q1 70.0 103.80
 CT-OS-HO 55.0 108.50 manually added from parm99.dat
 CI-OS-HO 55.0 108.50 manually modified from parm99.dat & frcmod.parmbsc0
 N*-CB-Q2 70.0000 134.500 QM_KOL0
 NB-CB-NA 70.0000 126.900 QM_KOL0
 CB-NA-HQ 50.0000 120.100 QM_KOL0
 CA-NA-HQ 50.0000 122.200 QM_KOL0
 CA-NA-CB 70.0000 117.900 QM_KOL0
 CB-Q2-NC 70.0000 114.300 QM_KOL0
 CB-CB-NA 70.0000 122.700 QM_KOL0
 CA-NC-Q2 70.0000 122.600 QM_KOL0
 O- Q2-NC 80.0000 121.600 QM_KOL0
 O- Q2-CB 80.0000 124.100 QM_KOL0
 Q2-CB-CB 63.0 119.20 parm10.dat

DIHE

X-Q1-N*-X 4 6.80 180.0 2.
 X-Q1-NB-X 2 20.00 180.0 2.
 OS-CT-N*-Q1 1 0.0649123 180.0 -4. chi KOL0
 OS-CT-N*-Q1 1 0.434356 180.0 -3. chi KOL0
 OS-CT-N*-Q1 1 0.952733 0.0 -2. chi KOL0
 OS-CT-N*-Q1 1 1.12182 180.0 1. chi KOL0
 CT-CT-N*-Q1 1 0.0797247 180.0 -4. chi KOL0
 CT-CT-N*-Q1 1 0.683783 180.0 -3. chi KOL0
 CT-CT-N*-Q1 1 0.686632 180.0 -2. chi KOL0

CT-CT-N*-Q1	1	0.808183	0.0	1.	chi KOL0
NB-CB-NA-HQ	1	4.150	180.0	2.	
CA-NA-CB-NB	1	4.150	180.0	2.	
CB-CB-NA-CA	1	4.150	180.0	2.	
CB-CB-NA-HQ	1	4.150	180.0	2.	
X-Q2-NC-X	2	8.00	180.0	2.	Parm10.dat
X-Q2-O-X	4	11.20	180.0	2.	Parm10.dat
X-Q2-CB-X	4	12.00	180.0	2.	Parm10.dat

IMPROPER

X-X-Q1-H5	1.1	180.	2.
CB-CT-N*-Q1	1.0	180.	2.

NONBON

Q1	1.9080	0.0860
Q2	1.9080	0.0860
HQ	0.6000	0.0157

AMBER parameter files for 7G-N1H,N3H,N2-imino

Developed at Institute of Bioorganic Chemistry PAS by Nivedita Dutta
MASS

Q1 12.01	0.360
Q2 12.01	0.616
HQ 1.008	0.161
ND 14.01	0.530
NP 14.01	0.530

BOND

Q1-H5 367.0	1.080
Q1-N* 440.0	1.371
Q1-NB 529.0	1.304
CQ-ND 481.0	1.340
NA-HQ 434.10	1.010
NP-HQ 434.10	1.010
Q2-O 570.0	1.229
Q2-NA 457.0	1.358
Q2-CB 447.0	1.419

NA-CB	461.0	1.3540
NP-CB	461.0	1.3540
H -ND	434.0	1.010
CA-ND	481.0	1.340
NP-CA	483.0	1.3390

MASS

Q1 12.01	0.360
Q2 12.01	0.616
HQ 1.008	0.161
ND 14.01	0.530
NP 14.01	0.530

BOND

Q1-H5	367.0	1.080
Q1-N*	440.0	1.371
Q1-NB	529.0	1.304
CQ-ND	481.0	1.340
NA-HQ	434.10	1.010
NP-HQ	434.10	1.010
Q2-O	570.0	1.229
Q2-NA	457.0	1.358
Q2-CB	447.0	1.419
NA-CB	461.0	1.3540
NP-CB	461.0	1.3540
H -ND	434.0	1.010
CA-ND	481.0	1.340
NP-CA	483.0	1.3390

DIHE

X -Q1-N*-X	4	6.80	180.0	2.
X -Q1-NB-X	2	20.00	180.0	2.
OS-CT-N*-Q1	1	0.0649123	180.0	-4. chi KOL0
OS-CT-N*-Q1	1	0.434356	180.0	-3. chi KOL0
OS-CT-N*-Q1	1	0.952733	0.0	-2. chi KOL0
OS-CT-N*-Q1	1	1.12182	180.0	1. chi KOL0
CT-CT-N*-Q1	1	0.0797247	180.0	-4. chi KOL0
CT-CT-N*-Q1	1	0.683783	180.0	-3. chi KOL0
CT-CT-N*-Q1	1	0.686632	180.0	-2. chi KOL0

CT-CT-N*-Q1	1	0.808183	0.0	1.	chi KOL0
X-CA-ND-X	4	9.60	180.0	2.	reinterpolated 93'
CA-NA-CB-NB	1	4.150	180.0	2.	
CA-NP-CB-NB	1	4.150	180.0	2.	
NB-CB-NA-HQ	1	4.150	180.0	2.	
NB-CB-NP-HQ	1	4.150	180.0	2.	
CB-CB-NA-CA	1	4.150	180.0	2.	
CB-CB-NP-CA	1	4.150	180.0	2.	
CB-CB-NA-HQ	1	4.150	180.0	2.	
CB-CB-NP-HQ	1	4.150	180.0	2.	
X-Q2-NA-X	2	8.00	180.0	2.	Parm10.dat
X-Q2-O-X	4	11.20	180.0	2.	Parm10.dat
X-Q2-CB-X	4	12.00	180.0	2.	Parm10.dat
X-NP-CA-X	4	1.5	180.0	2.	Parm10.dat

IMPROPER

X-X-Q1-H5	1.1	180.	2.	
CB-CT-N*-Q1	1.0	180.	2.	
X-X-ND-H	1.0	180.	2.	JCC,7,(1986),230
ND-NA-CA-NA	1.1	180.	2.	dac, 10/94
ND-NA-CA-NP	1.1	180.	2.	dac, 10/94

NONBON

Q1	1.9080	0.0860
Q2	1.9080	0.0860
HQ	0.6000	0.0157
ND	1.8240	0.1700
NP	1.8240	0.1700

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