

Supplementary information

tRNA^{Val} allows four-way decoding with unmodified uridine at the wobble position in *Lactobacillus casei*

Riko Sugita¹, Vincent Guérineau², David Touboul³, Satoko Yoshizawa⁴, Kazuyuki Takai¹, and Chie Tomikawa^{1*}

Author affiliations:

¹Department of Materials Science and Biotechnology, Graduate School of Science and Engineering, Ehime University, 3 Bunkyo-cho, Matsuyama, Ehime 790-8577, Japan

² Institut de Chimie des Substances Naturelles, CNRS UPR 2301, Université Paris-Saclay, 91198, Gif-sur-Yvette Cedex, France

³ Laboratoire de Chimie Moléculaire, CNRS UPR 9168, Ecole Polytechnique, IP-Paris, Route de Saclay, 91120, PALAISEAU, France

⁴ Université Paris-Saclay, ENS Paris-Saclay, CNRS UMR8113, Laboratory of Biology and Applied Pharmacology (LBPA), 91190, Gif-sur-Yvette, France

***To whom correspondence should be addressed:**

Chie Tomikawa

Department of Materials Science and Biotechnology, Graduate School of Science and Engineering, Ehime University, Bunkyo-cho 3, Matsuyama, Ehime 790-8577, Japan

Telephone: +81-89-927-9947

Email: tomikawa.chie.mm@ehime-u.ac.jp

Supplementary method

Nucleosides analysis of tRNA^{Val} by HPLC

Isolated tRNA^{Val} from *L. casei* was further purified by gel extraction. Nucleoside analysis of the purified tRNA^{Val} was performed according to our previous literature [1]. 4 µg of purified tRNA^{Val} was digested with 0.012 units of snake venom phosphodiesterase (Sigma), 20 µg of RNaseA (Roche), and 0.25 units of bacterial alkaline phosphatase BAP C75 (Takara) in 20 µl of 50 mM Tris-HCl (pH8.0) at 37°C overnight. The sample was purified using a PVDF centrifugal filter (Ultra free-MC-HV 0.45 µm, Millipore). Nucleosides were analyzed on an HPLC (Hitachi L-2000 system) equipped with a reverse phase C18 column (NUCLEOSIL 100 C18; 25 cm x 4.6 mm, 7 µm; GL Sciences, Inc). The solvent system consisted of buffer A (50 mM sodium phosphate, pH5.1) and buffer B (buffer A containing 70% methanol). The nucleosides (20 µl) were chromatographed using a flow rate of 1 ml/min with a multistep linear gradient as follows: 3% buffer B from 0-50min, 3-98% buffer B from 50-65 min, and 98-100% buffer B from 65-75 min. Standard pm⁶A was purchased from Sigma. The pm⁶A nucleotide was dephosphorylated with bacterial alkaline phosphatase before use. The elution points of 7-methylguanosine (m⁷G), 5-methyluridine (m⁵U), and pseudouridine (Ψ) were determined by enzymatic formations using the tRNA modification enzymes, TrmB, TrmA, and TruB, respectively.

Reference in Supplementary method

1. Tomikawa C, Yokogawa T., Kanai T., & Hori H. (2010) N7-Methylguanine at position 46 (m⁷G46) in tRNA from *Thermus thermophilus* is required for cell viability at high temperatures through a tRNA modification network. *Nucleic Acids Res* **38**, 942–957.