

Supplementary Material

Nucleobase carbonyl groups are poor Mg^{2+} inner-sphere binders but excellent monovalent ion binders—a critical PDB survey

Filip Leonarski^{1,2}, Luigi D'Ascenzo^{2,3}, and Pascal Auffinger^{2,*}

¹ Swiss Light Source, Paul Scherrer Institut, Villigen PSI, 5232, Switzerland

² Architecture et Réactivité de l'ARN, Université de Strasbourg, Institut de Biologie Moléculaire et Cellulaire du CNRS, Strasbourg, 67084, France

³ Department of Integrative Structural and Computational Biology, The Scripps Research Institute, La Jolla, CA 92037, USA

* To whom correspondence should be addressed. Tel: +33 388 41 70 49; Fax: +33 388 60 22 18;
Email: p.auffinger@ibmc-cnrs.unistra.fr

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About ribosomal PDB structures (resolutions ≤ 2.9 Å) and biological assembly choice:

The February 2017 PDB release contains 93 X-ray structures of ribosomes with resolutions ≤ 2.9 Å ([Table S1](#)) that comprise: (i) 7 *Thermus thermophilus* small subunits (30S), (ii) 1 *Deinococcus radiodurans* large subunit (50S), (iii) 44 *Haloarcula marismortui* large subunits (50S), (iv) 30 *Thermus thermophilus* 70S ribosomes, (v) 9 *Escherichia coli* 70S ribosomes and (vi) 2 *Saccharomyces cerevisiae* 80S ribosomes.

For the 41 ribosomal structures (70S and 80S) that are only available as mmCIF files, the biological assembly with the lowest average *B*-factor was retained for analysis (see [Table S1](#)). For the *T. thermophilus* 70S structures, numbering inconsistencies between the two or four assemblies resulting from in-house PDB annotations were found. When such discrepancies were noted, we chose to use the structure that has a numbering consistent with the 2D structures found at <http://apollo.chemistry.gatech.edu/RibosomeGallery> ([Petrov et al. 2013](#)) as noted in [Table S1](#).

Thermus thermophilus (30S)

In this ensemble of seven structures, **2VQE** has the best resolution (2.5 Å; [Table S1](#)). All structures contain $\text{Zn}^{2+}/\text{Mg}^{2+}$ and K^{+} has been assigned in six of them. For some of these structures, Ca^{2+} , Na^{+} , NH_4^{+} , Cl^{-} and acetate ions may also be present according to the crystallization conditions provided by the PDB. Both r-proteins S4 and S14 contain a zinc-finger domain ([Wimberly et al. 2000](#)).

Deinococcus radiodurans (50S)

5DM6 is given with a 2.9 Å resolution. Mg^{2+} is the only assigned ion. Zn^{2+} is not present since the zinc finger cysteine residues form disulfide bridges. Na^{+} , NH_4^{+} , spermidine and Cl^{-} may also be present according to the crystallization conditions noted in the PDB. But these ions were not assigned.

Haloarcula marismortui (50S)

In this ensemble of 44 structures, **1VQO** and **1VQ8** have the best resolutions (2.2 Å). Oddly, all these structures contain Cd^{2+} in replacement of Zn^{2+} . It was reported that the addition of 1 mM CdCl_2 was able to significantly improve the crystal growth of these 50S subunits ([von Bohlen et al. 1991](#)) —note that CdCl_2 has been used for crystallization of other ribosomal systems ([Wimberly et al. 1999](#); [Sigel et al. 2013](#)). The four L24e, L37e, L37ae, and L44e r-proteins contain zing finger motifs. Four of these domains, where Cd^{2+} replaces Zn^{2+} , were identified ([Klein et al. 2004](#)). In all these r-proteins, Cd^{2+} is tetrahedrally bound to four cysteine sulfur atoms. An additional weak and suspicious Cd^{2+} binding site is also reported. For some of these structures, NH_4^{+} and acetate ions may be present according to the crystallization conditions noted in the PDB. The **1FFK** (2.4 Å) crystallization conditions ([Ban et al. 2000](#)) indicate the use of following crystallization buffer: 1.2 M KCl, 0.5 M NH_4Cl , 100 mM KCH_3COO , 30 mM MgCl_2 , 7% PEG 6000, 15 mM tris, 15 mM MES, and 1 mM MgCl_2 at pH 7.1. The crystals were then stabilized by transfer to a solution containing 12% PEG 6000, 22% ethylene glycol, 1.7 M NaCl, 0.5 M NH_4Cl , 100 mM KCH_3COO , 30 mM MgCl_2 , and 1 mM CdCl_2 at pH 6.2.

This group of *H. marismortui* structures is very heterogeneous. In early structures, residue 1329:0 was wrongly assigned to an adenine as shown by $2F_o - F_c$ and $F_o - F_c$ maps ([Figure S12](#)). This error and other inaccuracies have been subsequently corrected in **4V9F** —a re-refinement of **1FFK** ([Gabdulkhakov et al. 2013](#))— and in the structures marked as such in [Table S1](#). The best

resolution structures with the corrected sequence are thus, **4V9F**, **1YHQ** and **3CC2** (2.4 Å). Modified nucleotides are assigned in **4V9F**. Other details regarding the modifications introduced into the *H. marismortui* refinement process can be found in ([Klein et al. 2004](#)).

Twenty-four of these ribosome structures contain Sr^{2+} . The use of SrCl_2 is related to the crystallization of antibiotic/ribosome complexes and was shown to improve crystal quality ([Zhang and Ferre-D'Amare 2014](#)). In one study, it is mentioned that crystals were soaked with antibiotic in a buffer containing: 1.7 M NaCl, 0.5 M NH_4Cl , 1 mM CdCl_2 , 100 mM KCH_3COO , 6.5 mM CH_3COOH and with either 30 mM MgCl_2 , or 21 mM MgCl_2 and 30 mM SrCl_2 , or no MgCl_2 and 100 mM SrCl_2 ([Blaha et al. 2008](#)). These conditions suggest that these ribosomes crystals include very high NaCl and NH_4Cl concentrations. The best resolution structure with Sr^{2+} and the corrected sequence is **3CCM** (2.55 Å).

Note that *H. marismortui* structures contain single ribosomal RNA chains, which are labeled in some structures “chain 0” and “chain A” in others. Here, these chains were considered equivalent.

***Thermus thermophilus* (70S)**

In this ensemble of 30 structures, **4Y4O** has the best resolution (2.3 Å). Zn^{2+} is present in 29 structures.

***Escherichia coli* (70S)**

In this ensemble of 9 structures, **4YBB** has the best resolution (2.1 Å as indicated in the PDB). It is also currently the best resolution of all ribosomal structures and contains a full array of modified nucleotides — note that the original publication mentions a 2.4 Å resolution ([Noeske et al. 2015](#)). For **4YBB**, a strong discrepancy between the *B*-factors of both assemblies present in the crystal unit must be noted (average *B*-factors around ≈ 67 and $\approx 123 \text{ Å}^2$ for assembly 1 and 2, respectively). As such, we limited our analysis to the first biological assembly. Besides 463 Mg^{2+} and 3 Zn^{2+} for the two assemblies, **4YBB** contains also acetate, 1,2-ethanediol, di(hydroxyethyl)ether, putrescine, spermidine, MPD, TRIS, tetraethylene- and pentaethylene-glycol. On the average, for all other *E. coli* ribosomes in [Table S1](#), one Zn^{2+} and 180 to 240 Mg^{2+} are present in each assembly.

***Saccharomyces cerevisiae* (80S)**

The best resolution structure is **4U4R** (2.8 Å). The two structures of this ensemble comprise each eight Zn^{2+} , $\approx 700 \text{ Mg}^{2+}$ and ≈ 500 osmium hexamine per biological assembly. Crystallization protocols are given in reference ([Ben-Shem et al. 2011](#)).

Crystallization buffers

To correctly analyze crystal structures, it is important to be informed about all the molecules present in purification and crystallization buffers. Some buffers frequently used for ribosome assembly and translation are polymix ([Jelenc and Kurland 1979](#)) and HiFi ([Gromadski and Rodnina 2004](#)).

- Polymix buffer: 5 mM $\text{Mg}(\text{OAc})_2$ or 14 mM $\text{Mg}(\text{OAc})_2$ and 95 mM KCl, 5 mM NH_4Cl , 8 mM putrescine, 1 mM spermidine, 1 mM DTT (2,3-dihydroxy-1,4-dithiobutane or dithioerythritol), 5 mM K_2HPO_4 , and 0.5 mM CaCl_2 .
- HiFi buffer: 3.5 mM MgCl_2 or 14 mM MgCl_2 and 30 mM KCl, 70 mM NH_4Cl , 8 mM putrescine, 0.5 mM spermidine, 1 mM DTT, and 50 mM Tris-HCl, pH 7.5.

Occupancy issues

Oddly, some structures have included Mg^{2+} ions with occupancies above 1.0 or very low B -factors that might hint to the presence of $\text{K}^+/\text{Ca}^{2+}/\text{Sr}^{2+}$ or transition metals and not to the presence of Mg^{2+} . See for example the Zn^{2+} ion suggested to be present in the RNA part of some ribosomes ([Leonarski et al. 2017](#)). Taking these ions into account would not be beneficial to our statistical analysis. Consequently, they were excluded from this survey. For instance, 1FJG and 1N32 display Mg^{2+} occupancies in the 0.22–1.47 range ([Carter et al. 2000](#); [Ogle et al. 2002](#)) and 4V9D, 4V51, 4U26, 4U27, 4U1V, 4V9P and 4V9O display water and Mg^{2+} with B -factors $\leq 1.0 \text{ \AA}^2$. In 4V9D, 262 out of 498 Mg^{2+} and 798 out of 802 waters have B -factors $\leq 1.0 \text{ \AA}^2$.

Table S1: List of PDB ribosomal structures with resolutions ≤ 2.9 Å. The best resolution structures in each category are highlighted in blue. In each category, structures are ordered according to PDB deposition date (first criterion), best resolution (second criterion) and best *R*-free (third criterion). The 5J5B and 5J7L structures were excluded given specific difficulties related to the authors very peculiar encoding of the associated mmCIF files. This table is an updated version of **Table S1** in reference ([Leonarski et al. 2017](#)) and comprises all structures with resolutions ≤ 2.9 Å instead of ≤ 3.0 Å.

PDB code	Deposited	Res. (<i>R</i> -Free)	Residue count	Ions	Reference
<i>T. Thermophilus (30S)</i> — 7 structures					
2UUB	03/2007	2.80 (0.24)	4086	Zn ²⁺ , Mg ²⁺ , K ⁺	(Weixlbaumer et al. 2007)
2UUA	03/2007	2.90 (0.25)	4086	Zn ²⁺ , Mg ²⁺ , K ⁺	(Weixlbaumer et al. 2007)
2UXC	03/2007	2.90 (0.26)	4086	Zn ²⁺ , Mg ²⁺ , K ⁺	(Dunham et al. 2007)
2VQE	03/2008	2.50 (0.28)	4086	Zn²⁺, Mg²⁺, K⁺	(Kurata et al. 2008)
2VQF	03/2008	2.90 (0.26)	4086	Zn ²⁺ , Mg ²⁺ , K ⁺	(Kurata et al. 2008)
3T1Y	07/2011	2.80 (0.27)	4065	Zn ²⁺ , Mg ²⁺	(Vendeix et al. 2012)
4B3M	07/2012	2.90 (0.25)	4072	Zn ²⁺ , Mg ²⁺ , K ⁺	(Perez-Fernandez et al. 2014)
<i>D. radiodurans (50S)</i> — 1 structure					
5DM6	09/2015	2.90 (0.27)	6490	Mg²⁺	(Kaminishi et al. 2015)
<i>H. marismortui (50S)</i> — with wrong adenine at position 1329 (see Figure S1) — 22 structures					
1FFK	07/2000	2.40 (0.26)	6825	Cd ²⁺ , Mg ²⁺ , K ⁺	(Ban et al. 2000)
1JJ2	07/2001	2.40 (0.22)	7279	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Klein et al. 2001)
1M90	07/2002	2.80 (0.22)	7282	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Hansen et al. 2002)
1QVG	08/2003	2.90 (0.26)	7288	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schmeing et al. 2003)
1S72	01/2004	2.40 (0.22)	7464	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Klein et al. 2004)
1VQO^a	12/2004	2.20 (0.25)	7478	Cd²⁺, Sr²⁺, Mg²⁺, K⁺, Na⁺, Cl⁻	(Schmeing et al. 2005a)
1VQ8^a	12/2004	2.20 (0.25)	7479	Cd²⁺, Sr²⁺, Mg²⁺, K⁺, Na⁺, Cl⁻	(Schmeing et al. 2005a)
1VQP	12/2004	2.25 (0.25)	7483	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schmeing et al. 2005a)
1VQK	12/2004	2.30 (0.25)	7480	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schmeing et al. 2005a)
1VQL	12/2004	2.30 (0.25)	7482	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schmeing et al. 2005a)
1VQM	12/2004	2.30 (0.25)	7482	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schmeing et al. 2005a)
1VQN	12/2004	2.40 (0.25)	7484	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schmeing et al. 2005b)
1VQ9	12/2004	2.40 (0.26)	7480	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schmeing et al. 2005a)
1VQ7	12/2004	2.50 (0.24)	7482	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schmeing et al. 2005b)
1VQ5	12/2004	2.60 (0.24)	7482	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schmeing et al. 2005a)
1VQ4	12/2004	2.70 (0.23)	7482	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schmeing et al. 2005a)
1VQ6	12/2004	2.70 (0.23)	7483	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schmeing et al. 2005b)
2OTL	02/2007	2.70 (0.25)	7462	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schroeder et al. 2007b)
2OTJ	02/2007	2.90 (0.24)	7463	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schroeder et al. 2007b)
2QEX	06/2007	2.90 (0.24)	7320	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Schroeder et al. 2007a)
3CPW	04/2008	2.70 (0.23)	7334	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Ippolito et al. 2008)
3OW2	09/2010	2.70 (0.25)	6801	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	—
<i>H. marismortui (50S)</i> — with correct guanine at position 1329 (see Leonarski et al. 2017) — 22 structures					
1YHQ	01/2005	2.40 (0.23)	7480	Cd²⁺, Sr²⁺, Mg²⁺, K⁺, Na⁺, Cl⁻	(Tu et al. 2005)
1YIJ	01/2005	2.60 (0.22)	7481	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Tu et al. 2005)
1YI2	01/2005	2.65 (0.21)	7481	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Tu et al. 2005)
1YIT	01/2005	2.80 (0.22)	7478	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Tu et al. 2005)
1YJ9	01/2005	2.80 (0.24)	7478	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Tu et al. 2005)
1YJW	01/2005	2.90 (0.22)	7481	Cd ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Tu et al. 2005)
3CC2	02/2008	2.40 (0.23)	7517	Cd²⁺, Mg²⁺, K⁺, Na⁺, Cl⁻	(Blaha et al. 2008)
3CCM	02/2008	2.55 (0.24)	7517	Cd²⁺, Sr²⁺, Mg²⁺, K⁺, Na⁺, Cl⁻	(Blaha et al. 2008)
3CC7	02/2008	2.70 (0.23)	7517	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Blaha et al. 2008)
3CCJ	02/2008	2.70 (0.23)	7517	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Blaha et al. 2008)
3CC4	02/2008	2.70 (0.24)	7517	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Blaha et al. 2008)
3CCE	02/2008	2.75 (0.23)	7517	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Blaha et al. 2008)
3CD6	02/2008	2.75 (0.24)	7520	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Blaha et al. 2008)
3CCU	02/2008	2.80 (0.22)	7517	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Blaha et al. 2008)
3CCL	02/2008	2.90 (0.22)	7517	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Blaha et al. 2008)
3CCV	02/2008	2.90 (0.22)	7517	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Blaha et al. 2008)
3CCQ	02/2008	2.90 (0.23)	7517	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Blaha et al. 2008)
3CMA	03/2008	2.80 (0.24)	7522	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Simonovic and Steitz 2008)
3G6E	02/2009	2.70 (0.23)	7217	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Gurel et al. 2009)
3G71	02/2009	2.85 (0.23)	7217	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Gurel et al. 2009)
3I56	07/2009	2.90 (0.24)	7217	Cd ²⁺ , Sr ²⁺ , Mg ²⁺ , K ⁺ , Na ⁺ , Cl ⁻	(Gurel et al. 2009)
4V9F^b	02/2012	2.40 (0.21)	7583^{c,d}	Cd²⁺, Mg²⁺, K⁺, Na⁺, Cl⁻	(Gabdulkhakov et al. 2013)

***T. thermophilus (70S)* — 30 structures**

4V51	07/2006	2.80 (0.31)	21886 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Selmer et al. 2006)
4V8I	12/2011	2.70 (0.25)	21484 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Polikanov et al. 2012)
4V9H	03/2013	2.86 (0.25)	11728 ^{c,d}	Mg ²⁺	(Tourigny et al. 2013)
1VY5	05/2014	2.55 (0.28)	21748 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Polikanov et al. 2014b)
1VY4	05/2014	2.60 (0.26)	21748 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Polikanov et al. 2014b)
1VY7	05/2014	2.80 (0.28)	21602 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Polikanov et al. 2014b)
1VY6	05/2014	2.90 (0.29)	21448 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Polikanov et al. 2014b)
4W2E	06/2014	2.90 (0.30)	11638 ^{c,d}	Zn ²⁺ , Mg ²⁺	(Gagnon et al. 2014)
4W2G	09/2014	2.55 (0.27)	21748 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Polikanov et al. 2014a)
4W2H	09/2014	2.70 (0.26)	21596 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Polikanov et al. 2014a)
4W2I	09/2014	2.70 (0.26)	21748 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Polikanov et al. 2014c)
4W2F	10/2014	2.40 (0.28)	21748 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Polikanov et al. 2014a)
4WPO	10/2014	2.80 (0.25)	24064 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Lin et al. 2015)
4WQF	10/2014	2.80 (0.26)	23898 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Lin et al. 2015)
4WQU	10/2014	2.80 (0.26)	23918 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Lin et al. 2015)
4WQY	10/2014	2.80 (0.27)	23760 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Lin et al. 2015)
4Y4O ^b	02/2015	2.30 (0.25)	21468 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Polikanov et al. 2015a)
4Y4P	02/2015	2.50 (0.28)	21748 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Seefeldt et al. 2016)
4Z3S	03/2015	2.65 (0.27)	21748 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Polikanov et al. 2015b)
4Z8C ^j	04/2015	2.90 (0.25)	21482 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Roy et al. 2015)
5DOY	09/2015	2.60 (0.28)	21754 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Polikanov et al. 2015b)
5FDV	12/2015	2.80 (0.24)	20986 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Seefeldt et al. 2016)
5F8K	12/2015	2.80 (0.29)	20948 ^{c,e}	Zn ²⁺ , Mg ²⁺ , K ⁺	(Seefeldt et al. 2016)
5FDU	12/2015	2.90 (0.23)	20974 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Seefeldt et al. 2016)
5HD1 ^j	01/2016	2.70 (0.27)	21484 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Gagnon et al. 2016b)
5HCQ ^j	01/2016	2.80 (0.25)	21472 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Gagnon et al. 2016b)
5HCR ^j	01/2016	2.80 (0.27)	21482 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Gagnon et al. 2016b)
5HCP ^j	01/2016	2.89 (0.27)	21474 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Gagnon et al. 2016b)
5J4B	03/2016	2.60 (0.26)	21748	Zn ²⁺ , Mg ²⁺ , K ⁺	(Melnikov et al. 2016)
5J4C	03/2016	2.80 (0.27)	21748	Zn ²⁺ , Mg ²⁺ , K ⁺	(Melnikov et al. 2016)
5J8B	04/2016	2.60 (0.26)	11953	Zn ²⁺ , Mg ²⁺	(Gagnon et al. 2016a)
<i>E. coli</i> (70S) — 9 structures					
4V9O	05/2013	2.90 (0.27)	45264 ^{c,f}	Zn ²⁺ , Mg ²⁺	(Pulk and Cate 2013)
4V9P	05/2013	2.90 (0.27)	44972 ^{c,f}	Zn ²⁺ , Mg ²⁺	(Pulk and Cate 2013)
4U27	06/2014	2.80 (0.26)	20808 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Noeske et al. 2014)
4U26	06/2014	2.80 (0.27)	20810 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Noeske et al. 2014)
4U20	06/2014	2.90 (0.28)	20794 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Noeske et al. 2014)
4U24	06/2014	2.90 (0.26)	20794 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Noeske et al. 2014)
4U25	06/2014	2.90 (0.26)	20794 ^{c,e}	Zn ²⁺ , Mg ²⁺	(Noeske et al. 2014)
4YBB ^b	02/2015	2.10 (0.23)	20744 ^{c,e,h}	Zn ²⁺ , Mg ²⁺	(Noeske et al. 2015)
5J5B ^b	04/2016	2.80 (0.20)	20745 ^{c,e,h}	Zn ²⁺ , Mg ²⁺	(Cocozaki et al. 2016)
<i>S. cerevisiae</i> (80S) — 2 structuresⁱ					
4U4R	07/2014	2.80 (0.25)	35344 ^{c,g}	Zn ²⁺ , Mg ²⁺	(Garreau de Loubresse et al. 2014)
4U3U	07/2014	2.90 (0.24)	35344 ^{c,g}	Zn ²⁺ , Mg ²⁺	(Garreau de Loubresse et al. 2014)

^a Although these ribosomes have the best 2.2 Å resolution for *H. marismortui*, they embed assignment errors as shown in **Figure S1** and should be considered with caution.

^b These structures include ribosomal modifications.

^c These very large ribosomal structures are only available in mmCIF format in the PDB.

^d These large structures contain one biological assembly (space group: P1211).

^e These large structures contain two biological assemblies (space group: P212121).

^f These large structures contain four biological assemblies (space group: P1211).

^g These large structures contain two biological assemblies (space group: P1211).

^h The *B*-factors of the first assembly are significantly higher than those of the second assembly that should consequently not be considered for further structural analysis.

ⁱ The numbering of these files are consistent with the 2D structures available at <http://apollo.chemistry.gatech.edu/RibosomeGallery> (Petrov et al. 2013).

^j The numbering of these files follows sequence and is not aligned towards *E. coli* ribosome.

^k Structure factors not available.

Table S2: Extract of the “ener_lib.cif” file (dated 04-04-1995/31-05-2000) showing coordination distance restraint values associated with Mg²⁺. It is advised to always check this and similar libraries before starting a structure refinement. Such files and their documented use should be included in the deposited PDB data. Note also the too long Mg²⁺ to oxygen coordination distance restraint values highlighted in yellow ([Leonarski et al. 2017](#)).

```
# -----
#      ener_lib.cif      4-APR-95/31-MAY-00
# -----
#
data_energy
#
# -----
#  _dictionary_name      ciflibdic
#  _dictionary_version    2.0
# -----
...

# --- BONDS ---
#
loop_
  _lib_bond.atom_type_1
  _lib_bond.atom_type_2
  _lib_bond.type
  _lib_bond.const
  _lib_bond.length
  _lib_bond.value_esd
#
#      _atom_type      atomic chemical type
#      _const          constant KBOND
#      _value          equilibrium length of this bond BOND0
#                      BOND - actual bond length
#
#                      ENERGY = KBOND * ( BOND - BOND0 )**2
#
#      Values for bond distances and sigmas are from (where it is possible):
#      International tables for crystallography
#      Volume C, 1992,
#      Edited by AJC Wilson
#      Published for IUCr by Kluwer Academic Publishers, Dordrecht/Boston/London
#      For carbon-carbon etc
#      Section: Typical Interatomic distances: Organic compounds
#      Authors: FH Allen, O. Kennard, DG Watson, L Brammer, AG Orpen and R Taylor
#      pages: 685-706
#
#      For metal radii and distances.
#      Section: Typical interatomic distances: Organometallic Compounds and
#      Coordination complexes of the d- and f-block metals
#      Authors: AG Orpen, L. Brammer, FH Allen, O Kennard, DG Watson and R Taylor
#      pages: 707-791
#
...
MG      .      metal      500.000  2.090      .      2.670      0.040
MG      NPA     metal      500.000  2.090      .
MG      NPB     metal      500.000  2.090      .
MG      OHA     metal      500.000  2.150      .      # ?
MG      OHB     metal      500.000  2.150      .      # ?
MG      OHC     metal      500.000  2.150      .      # ?
MG      O       metal      500.000  2.180      .      # ?
...
```

Table S3. MgRNA database Mg²⁺ binding sites involving O_r atoms and correspondence with present study. No “prior-knowledge” binding sites could be identified for these atoms suggesting that O_r atoms do generally not participate in Mg²⁺ binding sites. See also [Tables 2/S4](#) for Mg²⁺ binding sites centered on O_b and N_b atoms, respectively. With present state of structural knowledge, Mg²⁺ binding to O_r atoms seems only to occur with terminal nucleotides (see text and [Fig. S6](#)).

Order number	Potential binding sites ^a	MgRNA occurrence	Prior-knowledge (this study)
1.	O _r	22	<i>Poor stereochemistry</i>
2.	2O _r	6	<i>Poor occurrence and stereochemistry</i>
3.	O _r .O _b	1	<i>Poor occurrence and stereochemistry</i>
4.	2O _r .N _b	1	<i>Poor occurrence and stereochemistry</i>
5.	2O _r .2O _b	1	<i>Poor occurrence and stereochemistry</i>
6.	O _{ph} .O _r	49	<i>Poor stereochemistry</i>
7.	O _{ph} .2O _r	8	<i>Poor occurrence and stereochemistry</i>
8.	O _{ph} .O _r .2O _b	1	<i>Poor occurrence and stereochemistry</i>
9.	<i>cis</i> -2O _{ph} .O _r	36	<i>Poor stereochemistry</i>
10.	<i>cis</i> -2O _{ph} .2O _r	2	<i>Poor occurrence and stereochemistry</i>
11.	<i>cis</i> -2O _{ph} .O _r .O _b	2	<i>Poor occurrence and stereochemistry</i>
12.	<i>fac</i> -3O _{ph} .O _r	3	<i>Poor occurrence and stereochemistry</i>
13.	<i>mer</i> -3O _{ph} .O _r	19	<i>Poor stereochemistry</i>

^a The O_{ph} (OP1/2: anionic phosphate oxygen atoms), O_r (O2', O4': ribose oxygen atoms; O3', O5': phosphodiester oxygen atoms), O_b (O2, O4, O6: nucleobase carbonyl oxygen atoms) and N_b (essentially N7 purine nitrogen atoms) are derived from the MgRNA nomenclature ([Zheng et al. 2015](#)).

Table S4. MgRNA database Mg²⁺ binding sites involving N_b atoms and correspondence with data from a preceding study (Leonarski et al. 2017). Examples of potential “prior-knowledge” binding sites are highlighted in grey and reference to figures are given. See also Table 2/S3 for Mg²⁺ binding sites centered on O_b and O_r atoms, respectively.

Order number	Potential binding sites ^a	MgRNA occurrence	Prior-knowledge from (Leonarski et al. 2017)
1.	N _b	284	Figs 4A/4B
2.	O _r .O _b	1	Poor occurrence and stereochemistry —
3.	2N _b	158	Monovalent ion or transition metal binding site —
4.	2O _b .N _b	1	Poor occurrence and stereochemistry —
5.	O _b .2N _b	3	Poor occurrence and stereochemistry —
6.	2O _r .N _b	1	Poor occurrence and stereochemistry —
7.	O _{ph} .N _b	134	Fig. S8A (cis-) Fig. S8B (trans-)
7. ^b	O _{coo} .N _b	—	Not present in MgRNA Fig. 7B (trans-)
8.	cis-2O _{ph} .N _b	118	Fig. 7A/S7

^a The O_{ph} (OP1/2: anionic phosphate oxygen atoms), O_r (O2', O4': ribose oxygen atoms; O3', O5': phosphodiester oxygen atoms), O_b (O2, O4, O6: nucleobase carbonyl oxygen atoms) and N_b (essentially N7 purine nitrogen atoms) are derived from the MgRNA nomenclature (Zheng et al. 2015).

^b O_{coo} corresponds to the anionic oxygen atom of an Asp/Glu carboxyl/ate group and is not referenced by MgRNA.

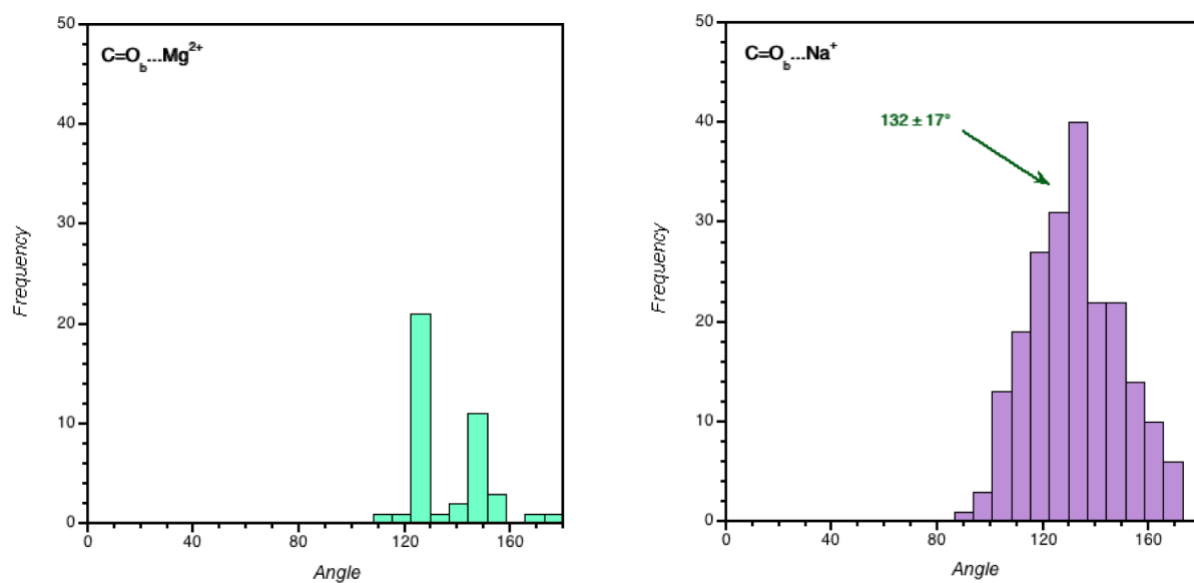


Figure S1. $C=O_b \dots Mg^{2+}/Na^{+}$ angle histograms derived from the CSD. (Left) $C=O_b \dots Mg^{2+}$ and (Right) $C=O_b \dots Na^{+}$ histograms. The average $C=O_b \dots Na^{+}$ angle is $132 \pm 17^\circ$. These histograms support the presence of the exclusion cone sketched [Fig. 2](#). Note that in the CSD, the structural environments are highly heterogeneous and the number of O_b -to- Mg^{2+} contacts is $\frac{1}{4}$ that of O_b -to- Na^{+} contacts.

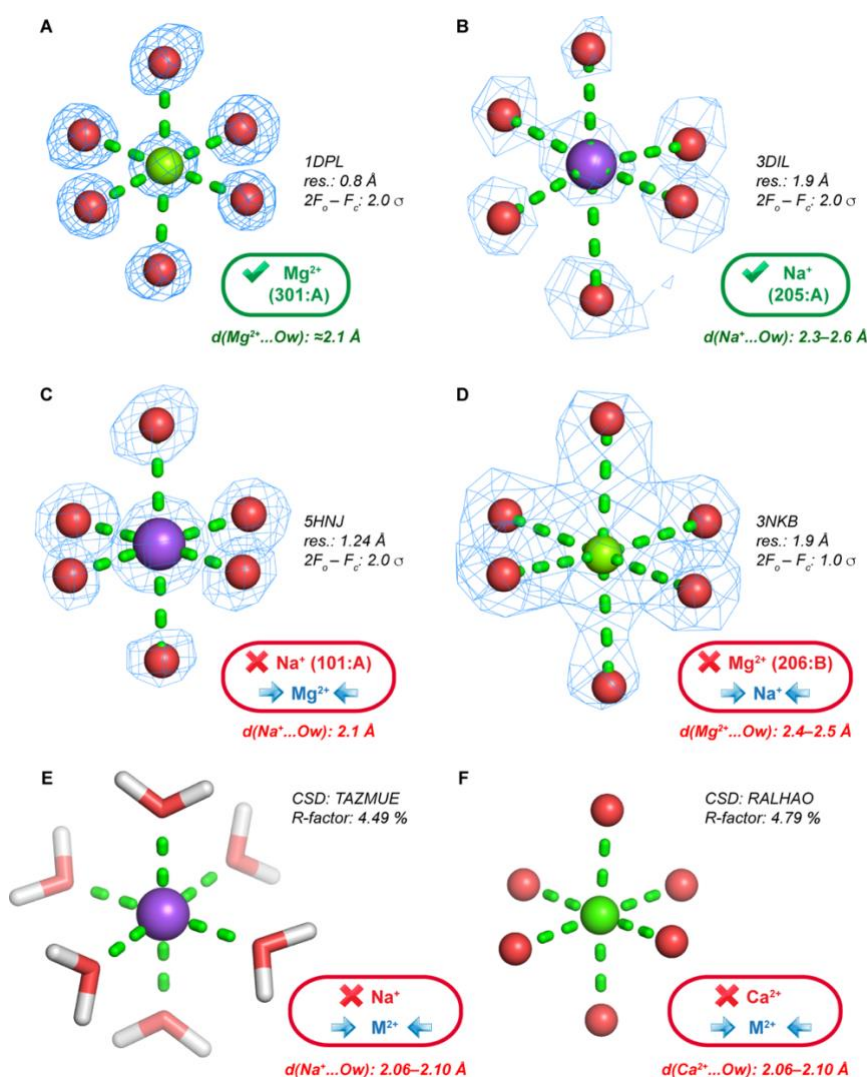


Figure S2. Stereochemically correct and unconvincing hexahydrated Mg²⁺/Na⁺/Ca²⁺ derived from PDB and CSD X-ray structures. (A) $d(\text{Mg}^{2+} \dots \text{Ow}) \approx 2.1 \text{ \AA}$ are compatible with Mg²⁺. (B) $d(\text{Na}^+ \dots \text{Ow}) \approx 2.4 \text{ \AA}$ are compatible with Na⁺. (C) $d(\text{Na}^+ \dots \text{Ow}) \approx 2.1 \text{ \AA}$ hint to Mg²⁺. (D) $d(\text{Mg}^{2+} \dots \text{Ow}) \approx 2.4 \text{ \AA}$ hint to Na⁺. (E) With respect to its $\approx 2.1 \text{ \AA}$ coordination distance d , this ion from the CSD should not be assigned to Na⁺. (F) Similarly, with respect to a $\approx 2.1 \text{ \AA}$ coordination distance characteristic of Mg²⁺, this ion should not be assigned to Ca²⁺.

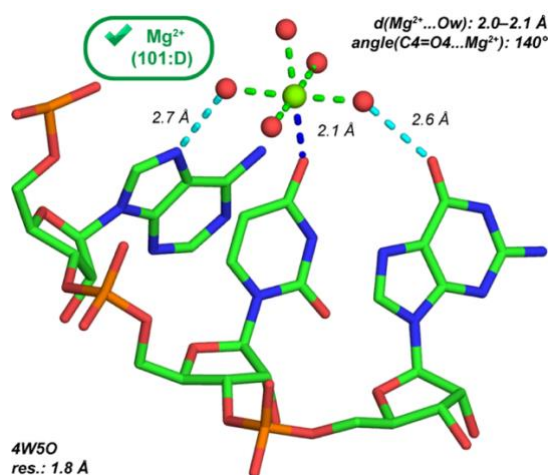


Figure S3. $\text{Mg}(\text{H}_2\text{O})_5^{2+}$ bound to the (U)O4 atom of a ApUpG sequence involves water mediated **contacts**. This coordination pattern, also observed in two other structures (PDBid: 4Z4D/E) is similar to that shown in [Fig. 3E](#) where the binding sequence is GpGpG.

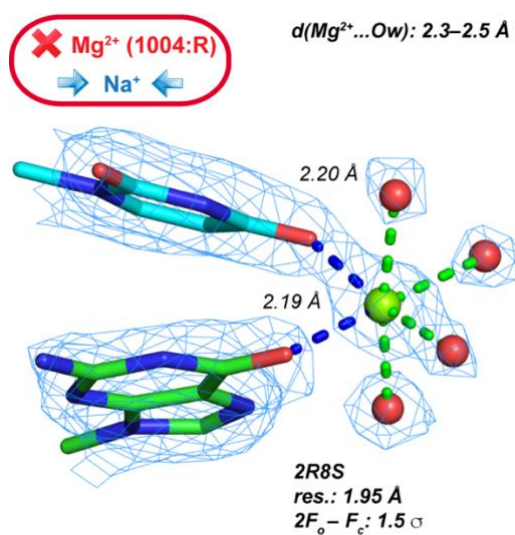


Figure S4. Unconvincing *cis*-2O_b Mg²⁺ binding site. Coordination distances to water hint to Na⁺. The ≈2.18 Å distances to the two O_b atoms point to the use of distance restraints — see [Fig. 6C](#) in ([Leonarski et al. 2017](#)) as well as [Table S2](#).

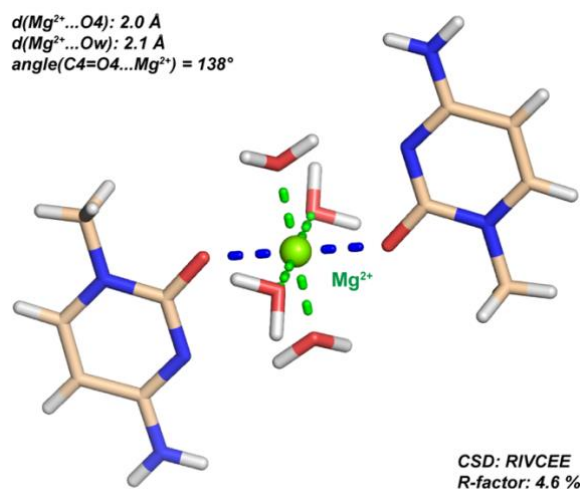


Figure S5. High-resolution CSD structure showing a *trans*-2O_b Mg²⁺ binding motif ([Gong et al. 2013](#)); see also ([Marino et al. 2016](#)). This motif is rare and has not been observed, with appropriate stereochemistry, in PDB structures. Note that the CSD crystallisation conditions usually largely differ from those used in biomolecular systems.

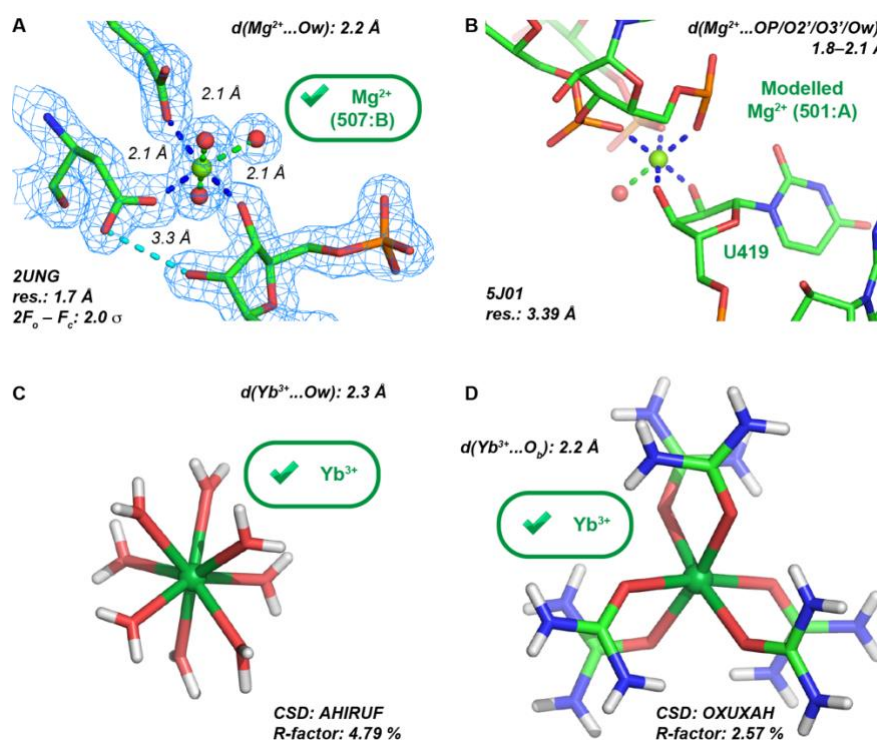


Figure S6. Mg²⁺ rarely binds to O2'/O3' hydroxyl groups; Yb³⁺ is rarely hexacoordinated. (A) From this unfrequent arrangement (2O_{coo}.O_r), it is suggested that the O3' atom coordinated to Mg²⁺ is deprotonated and may carry a negative charge ([Gan et al. 2008](#)). (B) A Mg²⁺ binding to the O2'/O3' atoms of a terminal residue (3O_{ph}.2O_r) modeled based on anomalous diffraction data of a group II intron Yb³⁺ derivative structure ([Costa et al. 2016](#)). (C) A Yb³⁺ with eight 1st shell water molecules commonly observed in the CSD ([Thuery 2009](#)). (D) A rare Yb³⁺ with six 1st shell oxygen atoms extracted from a CSD structure ([Lundberg et al. 2010](#)).

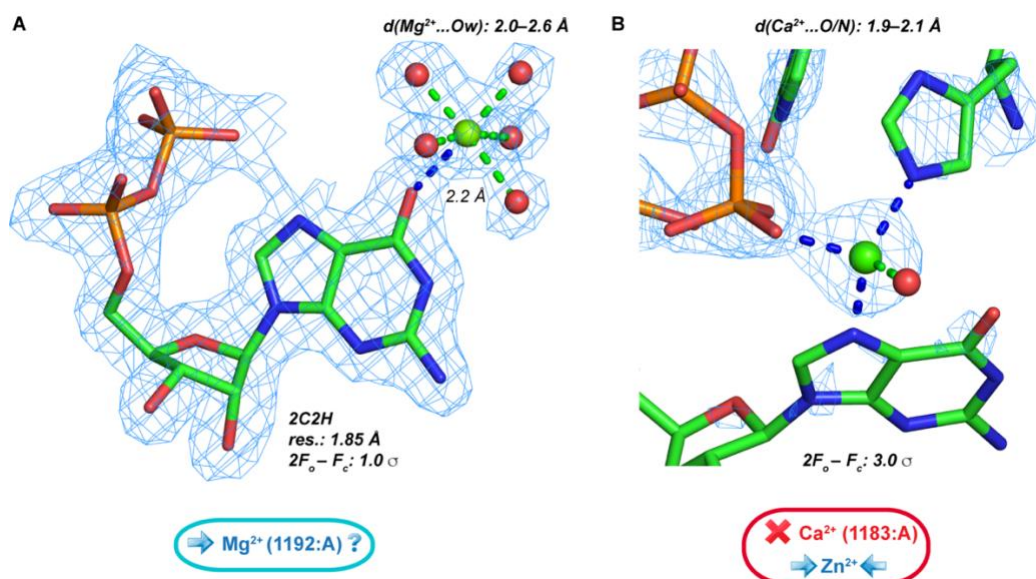


Figure S7. Unconvincing Mg^{2+} binding to a GDP containing DNA structure. (A) This Mg^{2+} binding to an O_b atom is unique among all PDB structures of nucleobase containing metabolites with resolution $\leq 2.9 \text{ \AA}$. (B) In the same structure, 12 tetrahedrally coordinated Ca^{2+} were assigned. This ion is more likely Zn^{2+} since Ca^{2+} has in all instances a coordination number ≥ 6 .

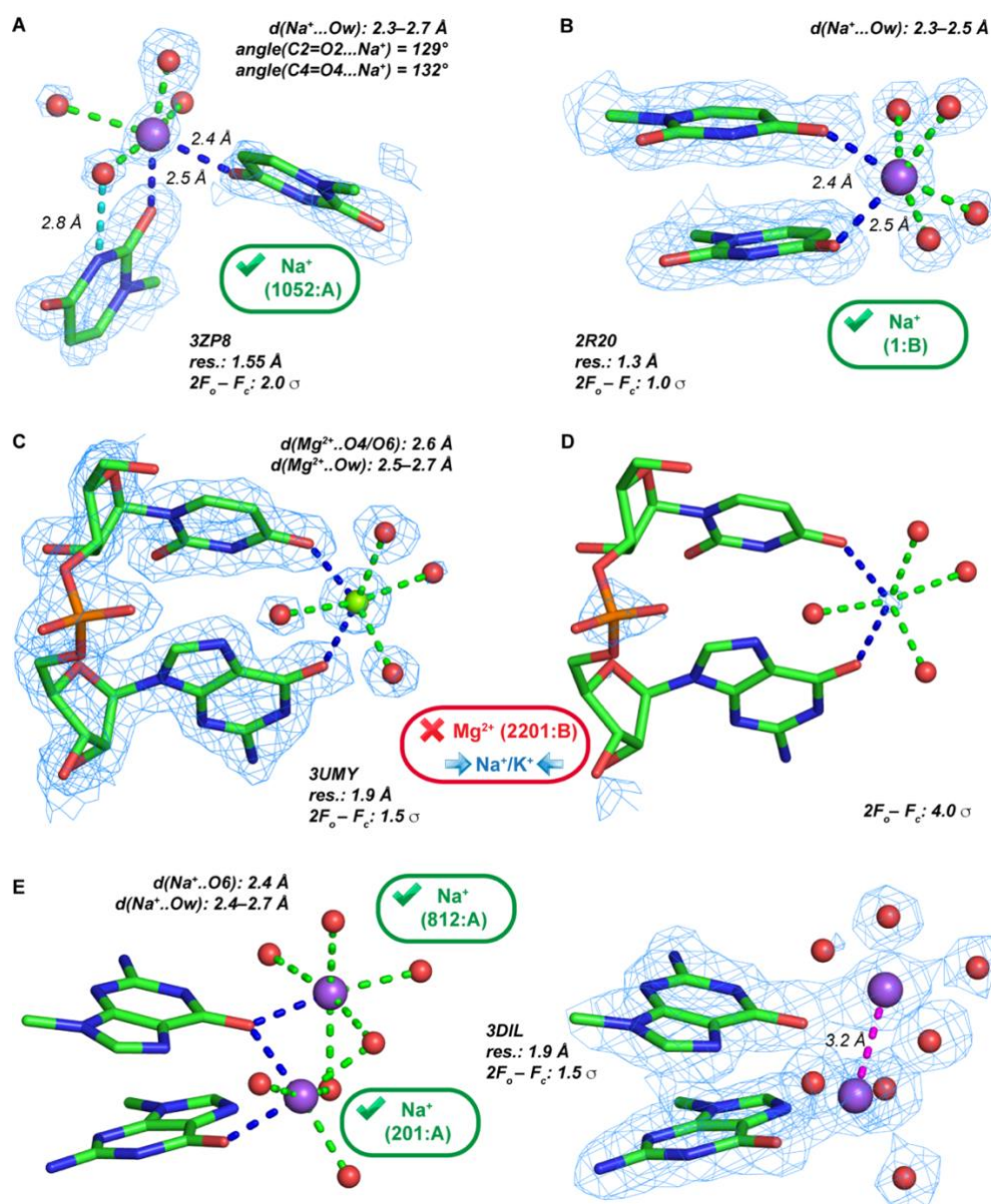


Figure S8. Na⁺ binding sites and a dimetallic Na⁺ cluster in PDB structures with resolution < 2.0 Å. (A) A *cis*-2O_b Na⁺ binding site resembling the *cis*-O_{ph}.O_b Mg²⁺ site shown Fig. 4B. (B) A frequent *cis*-2O_b Na⁺ site linking two consecutive uridines and resembling the *cis*-O_{ph}.O_b Mg²⁺ site shown in Fig. S4. (C) Improbable *cis*-2O_b Mg²⁺ binding site with coordination distances hinting to Na⁺ or K⁺. (D) The same binding site with densities shown at a 4.0 sigma level. The octahedral coordination suggests the presence of Na⁺ while both, the residual density at the 4.0 sigma level —ion not shown— and the longer coordination distances, suggests the presence of K⁺. In this case, anomalous diffraction data would be required for unambiguous ion assignment; see (Olieric et al. 2016). (E) Two views of a dimetallic Na⁺ cluster associated with a head-to-tail guanine arrangement similar to those described in (Leonarski et al. 2017).

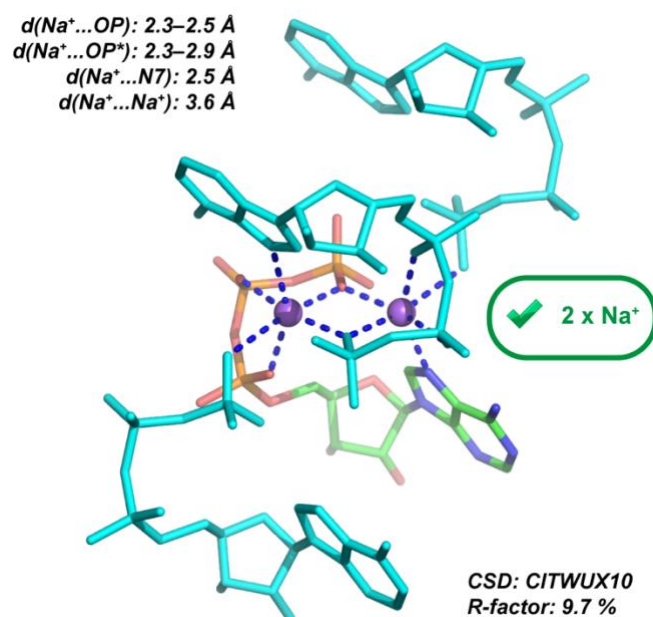


Figure S9. CSD structure of Na₂ATP with a dimetallic Na⁺ cluster ([Kennard et al. 1970](#); [Sugawara et al. 1991](#)). Each hexacoordinated Na⁺ is surrounded by five phosphate groups and one N7 atom. The ATP molecules in cyan are crystallographic symmetrics. OP* indicates coordination distances to a symmetric molecule.

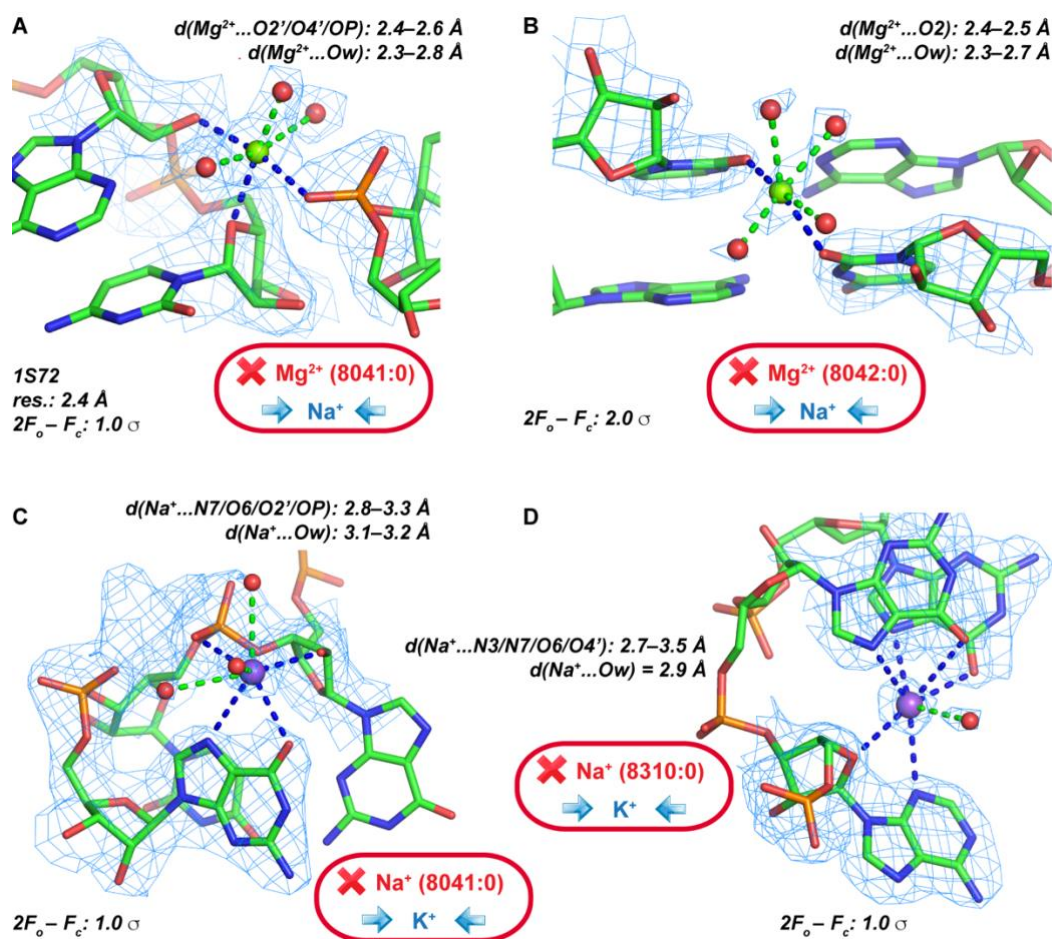


Figure S10. Unconvincing Mg^{2+} and Na^+ binding sites derived from *H. marismortui* 50S structure. (A/B) Coordination distances exclude Mg^{2+} and suggest the presence of Na^+ (C/D) Both, coordination distances and irregular coordination patterns exclude Mg^{2+} and point to the presence of K^+ .

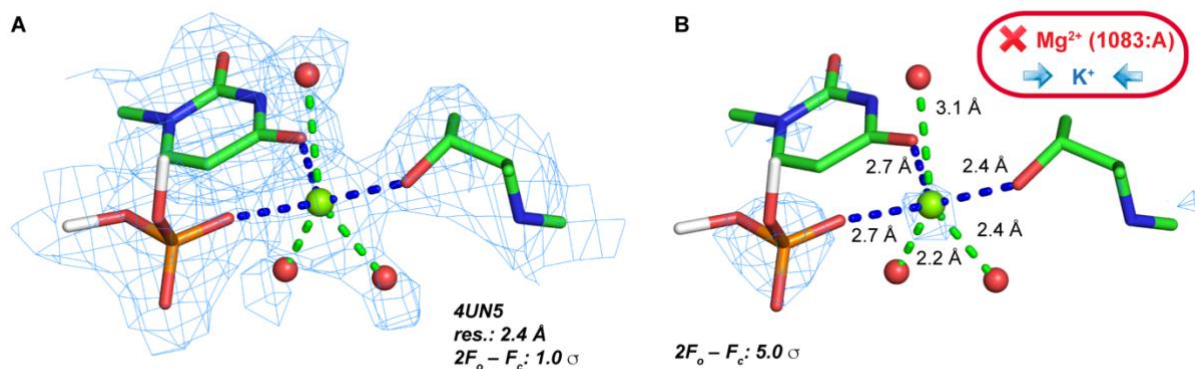


Figure S11. Unconvincing Mg²⁺ with a high sigma peak is probably K⁺ ([Anders et al. 2014](#)). **(A)** This hexacoordinated Mg²⁺ is bound to an OP, a (U)O4, a threonine hydroxyl group and three water molecules. Note that the two water molecules with short coordination distances are not placed at the center of the corresponding electron density patterns. **(B)** The Mg²⁺ density is still visible at 5.0 sigma. Therefore, this density pattern should rather be associated with K⁺ as suggested in ([Olieric et al. 2016](#)).

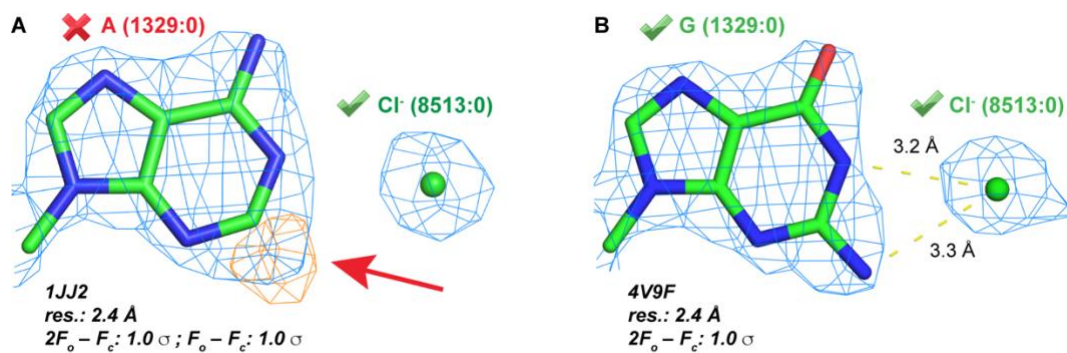


Figure S12. Nucleotide assignment error in early *H. marismortui* 50S structures. (A) Although a density for an amino N2 group is clearly visible in the $F_o - F_c$ map (red arrow), this nucleotide has been wrongly assigned to an adenine in 1FFK, 1JJ2 and related structures (Table S1) at odds with a Cl⁻ ion bound to its Watson-Crick edge (Auffinger et al. 2004; D'Ascenzo and Auffinger 2016). (B) This error has been corrected in 4V9F and related structures (Table S1).

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