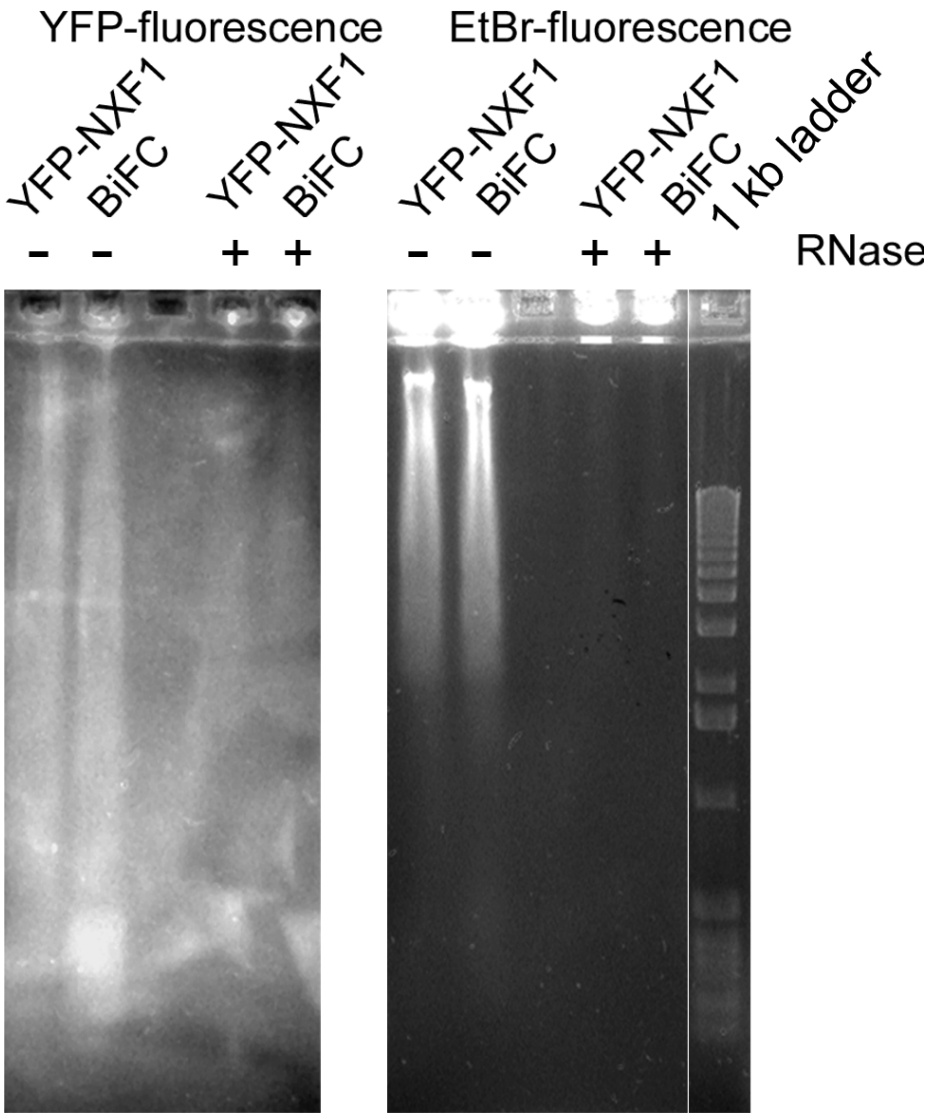

Supplementary Figure S1:

The soluble Y14/NXF1-BiFC and NXF1 fractions are composed of complexes of different size. The soluble supernatant of lyzed cells expressing Y14/NXF1-BiFC or YFP-NXF1 was separated on a 1% agarose gel and fluorescence was detected. (A) Y14/NXF1-BiFC- and YFP-NXF1 fluorescence is detected as a smear, indicating that the complexes are composed of different sizes (left). The smear is abolished upon RNase A treatment (right), confirming that RNA is a component of the observed smear. (B) Ethuidiumbromide (EtBr) staining of the same gel confirms the presence RNA before the RNase A treatment (left) and absence of RNA after the treatment (right).

Supplementary Methods:

Y14/NXF1-BiFC- or YFP-NXF1-expressing cells were lyzed on ice with PBS, supplemented with digitonin (50 μg/ml; Calbiochem), 1 mM EDTA and protease inhibitor cocktail (Roche). Cells were spun down at maximum speed for 20 minutes and the supernatant was saved. RNase A treatment was performed at 37°C for 20 minutes in lysis buffer supplemented with RNase A. Supernatants were loaded onto a vertical 1% agarose gel and migrated on ice. YFP fluoresescence was excited at 365 nm detected on a Biorad gel doc system. RNA was then stained with ethidiumbromide (Sigma) and detected on a Biorad gel doc system with the standard filters for excitation and emission for nucleic acid stained with ethidiumbromide. For details see Mazurkiewicz et al. (2006).

Supplementary Reference:

Mazurkiewicz, J., Kepert, J.F., Rippe, K. 2006. On the mechanism of nucleosome assembly by histone chaperone NAP1. *J Biol Chem* **281:**16462-16472.