Differential regulation of microRNA stability

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ABSTRACT
MicroRNAs (miRNAs) are endogenous single-stranded RNA molecules of about 21 nucleotides in length that are fundamental post-transcriptional regulators of gene expression. Although the transcriptional and processing events involved in the generation of miRNAs have been extensively studied, very little is known pertaining to components that regulate the stability of individual miRNAs. All RNAs have distinct inherent half-lives that dictate their level of accumulation and miRNAs would be expected to follow a similar principle. Here we demonstrate that although most miRNA appear to be stable, like mRNAs, miRNAs possess differential stability in human cells. In particular, we found that miR-382, a miRNA that contributes to HIV-1 provirus latency, is unstable in cells. To determine the region of miR-382 responsible for its rapid decay, we developed a cell-free system that recapitulated the observed cell-based-regulated miR-382 turnover. The system utilizes in vitro-processed mature miRNA derived from pre-miRNA and follows the decay of the processed miRNA. Using this system, we demonstrate that instability of miR-382 is driven by sequences outside its seed region and required the 3' terminal seven nucleotides where mutations in this region increased the stability of the RNA. Moreover, the exosome 3'→5' exoribonuclease complex was identified as the primary nuclease involved in miR-382 decay with a more modest contribution by the Xrn1 and no detectable contribution by Xrn2.

These studies provide evidence for an miRNA element essential for rapid miRNA decay and implicate the exosome in this process. The development of a biochemically amendable system to analyze the mechanism of differential miRNA stability provides an important step in efforts to regulate gene expression by modulating miRNA stability.

Keywords: miRNA stability; miRNA decay; miR-382; instability element

INTRODUCTION
MicroRNAs (miRNAs) are endogenous short noncoding single-stranded RNA molecules that post-transcriptionally regulate eukaryotic gene expression. While over 600 miRNAs have been identified in humans (Griffiths-Jones et al. 2008), as many as 1000 miRNAs have been predicted to be encoded by the human genome, representing ~3% of the total number of genes (Bentwich 2005).

Two cellular processing steps consecutively occur in the nucleus and the cytoplasm to generate mature miRNAs. First, primary miRNA (pri-miRNA) is cleaved into an approximate 70 nt-long hairpin precursor miRNA (pre-miRNA) by Drosha (Lee et al. 2003). Drosha, in conjunction with a regulatory subunit, DGCR8/Pasha, produces a pre-miRNA containing a 5' phosphate and a 3' hydroxyl with a 2-nt overhang recognition site for the second processing step (Denli et al. 2004; Gregory et al. 2004; Han et al. 2004; Landthaler et al. 2004). The pre-miRNA is exported to the cytoplasm by Exportin-5 (Zeng et al. 2003; Bohnsack et al. 2004; Lund et al. 2004), where a second processing step catalyzed by Dicer cleaves the pre-miRNA ~20-nt upstream of the 2-nt 3' overhang recognition site to produce a miRNA duplex intermediate. One of the two strands, generally the strand containing the least extent of hydrogen bonding at the 5' end of the duplex, becomes the mature miRNA, termed guide-strand, whereas the other strand is lost (Khvorova et al. 2003; Schwarz et al. 2003). Once selected, the mature miRNA is eventually incorporated into an RNA-induced silencing complex (RISC), and serves as the guide for RNA silencing (Murchison and Hannon 2004).
regarding the decay of miRNAs. By analogy, the regulation of mRNA stability plays a crucial part in the control of gene expression, where both the 5’ m7GpppN cap structure and the 3’ poly(A) tail contribute to the stability and translational competence of an mRNA (Furuichi and Shatkin 2000; Mangus et al. 2003). In contrast to their precursor pri-miRNAs, miRNAs as well as pre-miRNAs appear to have 5’ and 3’ unprotected ends that may render them accessible to exoribonucleases. However, in plants, miRNAs are protected from 3’–5’ exonucleolytic activity by a 2-O-methyl group or an oligo uridine tail added to their 3’ end (Li et al. 2005; Yu et al. 2005). Methylation or uridylation of the miRNA deters the activity of the recently identified 3’–5’ exoribonuclease SDN1 that specifically degrades single-stranded small RNAs in a sequence-independent manner (Ramachandran and Chen 2008). The SDN gene family is universally present in eukaryotes, but whether the human homolog of SDN1 exerts similar functions is currently unknown. In Caenorhabditis elegans, degradation of a subset of miRNAs was recently shown to be mediated by the nuclear 5’–3’ exoribonuclease, Xrn2 (Chatterjee and Grosshans 2009). Factors that can promote miRNA stability have also been reported recently. Translin, a DNA/RNA-binding protein, was shown to bind and increase the stability of miR-122 in vivo (Yu and Hecht 2008). Furthermore, post-transcriptional regulation of an miRNA is also dependent on both cis-elements and trans-acting factors. Although these cis-elements could occur anywhere on the miRNA, the majority of them are localized within the 3’ UTR (Decker and Parker 1993; Jackson 1993). Interestingly, a report by Hwang et al. (2007) showed that a hexanucleotide motif at the 3’ end of miR-29b was important for its regulation and was essential for its nuclear import.

We carried out a global analysis of miRNA levels following transcriptional shutoff in Human Embryonic Kidney (HEK293) cells and demonstrate that a majority of miRNAs does not appreciably decrease during the 8-h duration tested, with several exceptions. In particular, we identified miR-382, a miRNA involved in the inhibition of HIV-1 protein contributing to viral latency (Huang et al. 2007), as an unstable miRNA in cells. To further characterize the decay of miR-382, we developed an in vitro processing-coupled decay assay and identified a sequence within miR-382 that was essential for its reduced stability.

RESULTS

Differential stability of miRNAs

In order to analyze the extent to which miRNAs vary in their inherent stability, we used a microarray approach to specifically detect individual miRNA expression in HEK293 cells following a block in transcription. The miRNA microarray analysis was carried out with RNA extracted from cells treated with actinomycin D for 8 h to inhibit transcription. The efficiency of transcription inhibition was confirmed by RT-PCR of the unstable c-myc proto-oncogene. As expected, a significant reduction in c-myc mRNA levels were detected following 8 h of transcriptional shutoff, while the stable GAPDH mRNA was still abundantly present (Fig. 1A). Of the 310 human miRNA antisense probes contained on the NCode chip (Invitrogen), ~30% were expressed at appreciable levels in HEK293 cells and could be detected. Interestingly, 95% of the miRNAs remained stable and did not significantly change throughout the 8-h time course of the assay, reflecting half-lives greater than 8 h (Supplemental Fig. S1; Supplemental Table S1). Five miRNAs (miR-154, miR-224, miR-382, miR-204, and miR-211) were of significantly lower abundance at the 8-h actinomycin D time-point and appeared to have half-lives less than 8 h. However, it should be noted that since actinomycin D is expected to only affect the synthesis of pri-miRNA and not the subsequent processing events, microarray results represent the sum total of pri-miRNA stability and processing, post-miRNA stability and processing, and stability of the mature miRNA. These data indicate that the steady-state levels of only a subset of miRNAs decrease in HEK293 cells following transcriptional silencing for 8 h, while the levels of a majority of miRNAs do not change, as determined by microarray analysis.

In an effort to validate the microarray data, an independent real-time PCR approach was used to monitor levels of a subset of miRNAs before and after 8 h of actinomycin D-directed transcriptional shutoff. Six miRNAs were further tested and included two potentially unstable miRNAs (miR-224 and miR-382), two with apparent intermediate stability (miR-378 and miR-423), and two that appeared to be stable (miR-422b and miR-187) (Supplemental Fig. S1). Quantitative TaqMan real-time PCR that specifically detects and discriminates miRNA from their precursors was used. Consistent with the microarray results, miR-382 levels were reduced more than 50% after 8 h of actinomycin D treatment, while miR-423 was reduced by 20%, and the two apparently

![FIGURE 1](https://www.rnajournal.org/1033)

**FIGURE 1.** Differential stability of miRNAs in actinomycin D-treated HEK293 cells. (A) Total RNA from 8-h actinomycin D treated or untreated HEK293 cells were RT-PCR amplified with primers to detect the unstable c-myc proto-oncogene mRNA or stable GAPDH mRNA to confirm transcriptional inhibition. (B) Microarray results for a subset of miRNAs were verified by quantitative RT-PCR using TaqMan miRNA assay kit (Applied Biosystems). Data represent the ratio of miRNA at 8 h actinomycin D treatment relative to untreated cells and represent the mean of at least three independent experiments ± standard errors.
stable miRNAs, miR-422b and miR-187, remained unchanged (Fig. 1B). In contrast to the microarray results, the levels of miR-224 and miR-378 remained unchanged. The greater sensitivity of the real-time PCR approach suggests that these results more accurately represent the cellular miRNA levels and indicate that miR-224 and miR-378 are stable in HEK293 cells, while miR-382 appears to be selectively and reproducibly unstable in HEK293 cells.

**miR-382 is unstable in cells**

To more directly follow the stability of miRNAs, a Dicer knockdown cell line was used to curtail pre-miRNA processing and enable a more focused analysis of miRNA stability minimally affected by the extent of pre-miRNA processing. A HEK293 cell line containing a tetracycline-regulated short hairpin RNA (shRNA) directed against Dicer, which can efficiently reduce Dicer levels following 48 h of induction, was used (Schmitter et al. 2006). The fate of four miRNAs, miR-382, miR-187, miR-224, and miR-378, was followed over 3 d with quantitative real-time PCR following induction of Dicer-directed shRNA. The levels of each miRNA tested are presented relative to the stable miR-378. As shown in Figure 2, the relative levels of each miRNA are distinctly affected, demonstrating differential stability of miRNAs. Consistent with the above data, miR-382 levels were the most prominently reduced, with miR-187 and miR-224 displaying intermediate profiles compared with miR-378. Collectively, our data demonstrated that miR-382 was consistently unstable relative to miR-187, miR-224, and miR-378 in HEK293 cells.

To identify the exoribonucleases that affect miR-382 levels, a lentiviral shRNA delivery system was implemented to individually knockdown the exoribonucleases, Xrn1, Xrn2, or the exosome complex. The efficiency of the knockdowns is shown in Figure 3A, where an 85% and 83% reduction in the mRNAs encoding the Xrn1 and Xrn2 3′–5′ exoribonucleases was obtained, respectively. A 78% reduction in any of the three nucleases (Fig. 3B). Collectively, these data support a role for both the exosome and Xrn1 in the selective decay of miR-382, with the exosome appearing to fulfill a more primary role.

**Development of an in vitro miRNA decay system**

To enable a mechanistic understanding of components that govern miRNA stability, we initiated studies to analyze miRNA stability in vitro. We initially adapted an assay that we have successfully used to study mRNA decay (Wang et al. 1999) and followed the decay of the stable miR-378 and unstable miR-382. However, when using either single-stranded or double-stranded miRNA to follow their decay in extract, differential stability between miR-382 and miR-378 was not detected (Fig. 4). These data suggest that the distinct stabilities between the two miRNAs observed in cells might be due to specific ribonucleoprotein complexes that normally form on the miRNA during pre-miRNA processing, which were not present on the naked RNA added to the reaction.

To overcome this limitation, we rationalized that miRNAs that were generated in vitro from pre-miRNA would more closely mimic endogenous conditions. Pre-miRNA processing might promote assembly of endogenous ribonucleoprotein complexes onto the miRNA. A coupled in vitro pre-miRNA processing and miRNA decay assay was
The miRNA derived from pre-miRNA processing can be mimicked within a cell-free assay system that utilizes exogenous Dicer and a shift of the reaction to 18°C to facilitate more efficient pre-miRNA processing (Zhang et al. 2002). The decay assay was initiated by the addition of cell extract lacking exogenous Dicer and a shift of the reaction to 18°C to minimize pre-miRNA processing. Consistent with the data observed in cells, miR-378 was reproducibly more stable relative to miR-382 with half-lives of >160 and 80 min, respectively (Fig. 5A,B; summarized in Fig. 5C). These data demonstrate that the 3′ terminal seven nucleotides of miR-382 (positions 16-22) are necessary for its rapid decay.

The essential region within the second half of miR-382 was determined with three additional mutations to further refine the minimal sequence necessary for instability (Fig. 6A,C). Substitutions at positions 13–15 (miR-382mut13-15) had no significant effect on the stability of the miRNA, indicating that these nucleotides do not contribute to the decay of miR-382. Interestingly, two sets of mutations that disrupted sequences within the 3′ terminal seven nucleotides (miR-382mut16-18 and miR-382mut19-22) both increased stability of the miRNA. These data demonstrate that the 3′ terminal seven nucleotides of miR-382 (positions 16–22) are necessary for the observed rapid decay of this miRNA.

DISCUSSION

Despite an extensive analysis of the multiple processes involved in the generation of miRNAs, relatively little is known regarding the decay of miRNAs and the contribution of miRNA stability to their accumulation in a cell. In this study, we demonstrate that miRNAs can possess intrinsic distinct stabilities. Using a global analysis approach to identify miRNAs with reduced levels following a transcriptional shutoff, we identified a subset of miRNAs whose steady-state levels decreased significantly. In particular, miR-382 was found to be unstable in HEK293 cells, and the reduced stability was dependent on the 3′ terminal seven nucleotides of the miRNA. Our data demonstrate that a specific sequence within a miRNA could dictate its stability, and suggests that other miRNAs may also have intrinsic elements that modulate their stability.

Our global analysis of miRNA levels following transcriptional shutoff indicated that the miRNA levels for a majority of miRNAs detected did not appreciably change during the 8-h transcriptional arrest (Supplemental Fig. S1). However, it should be noted that although transcription was terminated, processing of pri- and pre-miRNAs into miRNAs is expected to continue under these conditions. If the processing of a particular miRNA precursor is regulated and the processing occurs relatively slowly, miRNAs could be replenished and appear to be more stable than they are. In addition, whether actinomycin D, which has been shown to aberrantly stabilize some miRNAs (Shyu et al. 1989), could also impact miRNA stability is currently unknown. Therefore, it is possible that the observed ~95% of the miRNAs that did not decrease in overall abundance could be an overestimation.
enriched miRNAs following surprisingly, a recent report detected the levels of five brain-analyses only a subset of miRNAs appear to be unstable, decay rates relative to the stable miR-378. Although in our distinct relative decay kinetics, with miR-382 as the most under these conditions, all four miRNAs tested displayed mediated our hypothesis that miRNAs possess distinct stabilities.

The development of an in vitro–coupled pre-miRNA processing and miRNA decay assay that recapitulates the differential stabilities of miR-378 and miR-382 observed in cells (Tang et al. 2008). However, it is likely that the 3′ terminus of the miRNA could be accessible to factors, regardless of their association with RISC (see below). In addition, components of the RISC may also influence the stability of miRNAs. A central component of RISC, the Ago2 protein (Filipowicz 2005), was shown to have an RNase-independent function of post-transcriptionally enhancing expres-

FIGURE 5. Coupled pre-miRNA processing and miRNA decay in vitro. Coupled processing and decay assays were carried out with 5′-end 32P-labeled pre-miR-382 (A) and pre-miR-378 (B). 32P-labeled pre-miRNAs were incubated with 10 μg of Dicer-overexpressing HEK293 cytoplasmic S15 extract at 37°C for 30 min to process the pre-miRNA into mature miRNAs. Decay of the mature miRNA was then followed by the addition of 50 μg of HEK293T cytoplasmic S15 at 18°C (lanes 2–5). Lanes 6–8 show the extent of background processing that occurs during the decay assay. The 32P-labeled pre-miRNA in lanes 6–8 were incubated directly at 18°C with a mixture of 10 μg of cytoplasmic S15 extract derived from Dicer-overexpressing cells and 50 μg of cytoplasmic S15 extract from HEK293 cells. Reactions were terminated as in the legend to Figure 4. The pre-miRNA, mature miRNA, and loading controls are indicated on the right and the size markers on the left. (C) Graphic representation of miR-382 and miR-378 decay relative to the level of miRNA present in the ‘0’ time point are shown. 32P-signals were quantified using ImageQuant software. Data represent the mean of at least three independent experiments ± standard errors.

Dicer knockdown conditions, which minimize pre-miRNA processing and accentuate miRNA turnover, further substantiated our hypothesis that miRNAs possess distinct stabilities. Under these conditions, all four miRNAs tested displayed distinct relative decay kinetics, with miR-382 as the most unstable, miR-187 and miR-224 displaying intermediate decay rates relative to the stable miR-378. Although in our analyses only a subset of miRNAs appear to be unstable, surprisingly, a recent report detected the levels of five brain-enriched miRNAs following α-amanitin-directed transcriptional shutoff, and found that all of the five miRNAs decreased rapidly with <50% of the detected miRNAs remaining by 3 h (Sethi and Lukiw 2009). Whether the rapid decrease in these five miRNAs is a general property of neuronal cells or serendipitous, is currently unknown.

Further support for selective post-transcriptional regulation of miRNAs was recently reported. The testis-brain RNA-binding protein, Translin, was shown to stabilize miR-122 when overexpressed in HUH-7 cells and selectively bind this miRNA in vitro (Yu and Hecht 2008). Most miRNAs are tightly associated with the RISC and only a small fraction are thought to be independent of RISC in

Bail et al.
vivo provides a biochemical system to enable mechanistic analysis of miRNA stability and potentially other noncoding RNA stabilities. Using this system, we found that the last seven nucleotides of miR-382 are essential for conferring instability onto the miRNA. Similarly, a hexanucleotide motif contained at the 3' end of miR-29b has also been implicated in miRNA regulation by directing enrichment of this miRNA into the nucleus (Hwang et al. 2007). The positioning of the required region for stability and transport within miR-382 and miR-29b, respectively, is particularly intriguing. The 5' region of a given miRNA, referred to as the seed region, is important for directing a miRNA to its mRNA target. Structural analysis of a bacterial argonaute protein in complex with a guide DNA and target mRNA revealed that the 3' end of a DNA-guide strand (residues 12–19) displayed fewer contacts with the protein than its extensively anchored 5' seed counterpart (Wang et al. 2008), indicating its potential accessibility to proteins. Positioning of the miRNA regulatory region to the 3' end and likely outside the mRNA and RISC interaction regions could render the miRNA more accessible to trans-factors that can exert an influence onto the miRNA, perhaps analogous to the function of the 3'-untranslated region of an mRNA. Future studies to identify trans-factors that can function through the miR-382 3'-end element will begin to address these possibilities.

Although currently underappreciated, regulation of miRNA stability is likely to emerge as an important step in the control of miRNA levels and, consequently, a broader range of regulation that can impact gene expression. Modulation of miR-382 can have significant biological relevance, as is illustrated by its contribution to differential HIV-1 protein production in resting and activated CD4+ T cells, as well as modulation of HIV-1 latency (Huang et al. 2007). The resistant nature of latent HIV-1 to therapeutic intervention (Lassen et al. 2004; Peterson et al. 2007) indicates that efforts aimed at preventing latency would significantly benefit therapeutic efficacy and one avenue could involve controlling stability of miR-382.

Having developed a coupled premiRNA processing and miRNA stability assay can now provide an appropriate platform to identify factors conferring miRNA stability and to mechanistically decipher their molecular function. The demonstration that miRNAs could possess distinct intrinsic stabilities will facilitate new areas of investigation into miRNA regulation. Moreover, an understanding of how gene expression can be controlled by miRNA stability may provide novel therapeutic applications to manipulate the expression of a broad spectrum of genes associated with human disorders.

MATERIALS AND METHODS

Plasmid generation

The Dicer coding region was excised from the pcDNA3-T7-Dicer plasmid (kindly provided by J. Liu, Sloan-Kettering Institute) with HindIII and XhoI and inserted into the same sites in pcDNA3-FLAG plasmid to generate pcDNA3-Flag-Dicer, which expresses FLAG-tagged Dicer protein.

Cell culture, transfections, and extract preparation

HEK293 and HEK293T cells were maintained in Dulbecco’s Modified Eagle Media (DMEM, Invitrogen) supplemented with 10% fetal bovine serum and a mixture of antibiotic–antimycotic. HEK293T cells were transfected at 80% confluency with pcDNA3-FLAG-Dicer using Lipofectamine 2000 (Invitrogen) as described by the manufacturer. Twenty-four hours post-transfection, cells

**FIGURE 6.** Requirement of miR-382 3' terminus for its rapid decay. (A) Processing-coupled in vitro decay assays were carried out with pre-miR-382 containing substitutions within the first or second half of the miRNA or (B) at positions 13–15, 16–18, and 19–22 in the second half of the miRNA. Reaction conditions and labeling are as described in the legend to Figure 4. (C) The sequence of wild-type and mutants miR-382 are shown at the left, where lines demarcate the substituted sequences. Bars represent the ratio of wild-type and mutant miR-382 levels expressed as a ratio of miRNA remaining after an 80-min decay reaction relative to the initial level of miRNA prior to the decay. Data represent the mean of at least three independent experiments ± standard errors. Significance was determined by ANOVA using Graph Pad InStat software. *P < 0.05, **P < 0.01, ***P < 0.001.
were harvested and cytoplasmic S15 extract isolated as previously described (Song and Kiledjian 2007). HEK293 cells at 80% confluence were treated with actinomycin D (5 μg/mL, Sigma) for 8 h to inhibit transcription. Knockdown of Dicer in HEK293 conditional Dicer knockdown 2b2 cells was carried out as previously described (Schmitter et al. 2006). Knockdowns of Xrn1, Xrn2, and Rrp41 were obtained in HEK293T by lentiviral RNAi-directed gene silencing and selection with puromycin (3 μg/mL) as described by the manufacturer (Sigma).

**RNA isolation and synthesis**

Total RNA isolations were carried out with TRIzol (Invitrogen). miRNAs were enriched from total RNAs using two Microcon columns following the protocol described by the manufacturer (Millipore) to enrich for low-molecular weight RNAs (LMW). The efficiency of actinomycin D-mediated transcriptional shutoff was verified by RT-PCR of the proto-oncogene c-myc mRNA, which has a half-life less than 1 h in mammalian cells. Briefly, total RNAs were treated with Turbo DNase (Ambion) to degrade DNA prior to oligo(dt) (IDT)-directed reverse transcription with M-MLV-RT (Promega) according to the manufacturer’s instructions. Amplification of the cDNA was carried out by PCR at 94°C for 30 sec, 60°C for 20 sec, and 72°C for 30 sec, for 22 cycles using Taq DNA polymerase.

DNA templates to generate pre-miRNAs were obtained by PCR of 60 base oligonucleotide pre-miRNA templates using 5’ SP6 and 3’ pre-miRNA specific primers (Supplemental Table S2) by Advantage Taq DNA polymerase (Clontech) according to the manufacturer. Pre-miRNAs were transcribed with SP6 RNA polymerase (Promega) according to the manufacturer. The SP6’ end of the resulting RNA was dephosphorylated with calf intestine phosphatase (New England Biolabs). For miRNA decay assays, pre-miRNAs were transcribed with 5’-32P-labeled ATP (Perkin Elmer) using T4 polynucleotide kinase (New England BioLabs).

**Genome-wide analysis of miRNA stability**

The LMW RNAs (500 ng) were subsequently tagged using the 3DNA Array 900 miRNA direct protocol from Genisphere and used to probe a miRNA microarray chip as follows. Briefly, a poly(A) tail was first added to the 3’ hydroxyl end of the LMW RNAs. The LMW RNAs were subsequently ligated to a capture sequence tag via a bridging oligonucleotide. The capture sequence is a 31-nt-long oligonucleotide partially complementary to the bridging oligo and attached to a 3DNA dendrimer labeled with either Cy3 or Cy5 fluorochromes. The 19-nt-long bridging oligonucleotide consists of 9 nt that are complementary to the capture sequence tag and 10 nt complementary to the added poly(A) tail. Sequence-tagged LMW RNAs were hybridized to the NCode miRNA microarrays (Invitrogen) containing antisense miRNA probes according to Goff et al. (2005). A set of 10 synthetic control miRNAs (NCode Control Set, Invitrogen) are spiked into the microarray labeling reaction to minimize and monitor experimental variability between time points and across microarrays. These probes do not cross-react with any probes for endogenous miRNAs on the microarray.

Arrays were scanned using an Axon GenePix 4000B scanner (Molecular Devices) and median spot intensities collected using Axon GenePix 4.0 (Molecular Devices). Data analysis and manipulation were conducted in GeneSpring 7.0 (Agilent) using quantile analysis (with the R package limma; http://www.bioconductor.org). Results were normalized to the spike-in NCode control probe signals to correct for global changes in miRNA populations.

Real-time PCR confirmation was carried out with Turbo DNase (Ambion) treated total RNA and RT-PCR using the appropriate TaqMan microRNA assay (Applied Biosystems) as described by the manufacturer.

**Coupled pre-miRNA processing and miRNA decay**

Coupled pre-miRNA processing and miRNA decay assays were carried out with 5’-end labeled in vitro synthesized pre-miR-382 and its derivatives or purchased pre-miR-378 (Dharmacon) (Supplemental Table S2). Pre-miRNA hairpins were formed by incubation at 95°C for 2 min and slow cooled to room temperature in annealing buffer (20 mM KAc, 6 mM HEPES at pH 7.4, 0.4 mM MgOAc). Pre-miRNA processing reactions were carried out with 1000 cpm of 32P-labeled pre-miRNA incubated at 37°C for 30 min in JVD-1 buffer (20 mM Tris at pH 7.5, 200 mM KAc, 4 mM MgOAc, 4 mM DTT, 20 mM creatine phosphate, 2 mM ATP, 0.8 mM GTP, 0.2 mM spermine) containing 10 μg of cytoplasmic extract from Dicer-overexpressing HEK293T cells pretreated for 10 min with 10 ng proteinase K (Sigma) to activate Dicer activity (Zhang et al. 2002). The decay reactions were subsequently initiated by the addition of 50 μg of HEK293T cytoplasmic extract and followed at 18°C for the indicated times. Control reactions to determine the extent of background processing during the decay reactions were determined by incubation of pre-miRNA with both proteinase K-pretreated Dicer-overexpressing and HEK293T cytoplasmic extracts at 18°C for the same time intervals. Reactions were stopped by addition of 150 μL of ULB (10 mM Tris at pH 7.5, 0.35 M NaCl, 2% SDS, 10 mM EDTA, 7 M Urea) spiked with 100 cpm 5’-end 32P-labeled loading control, phenol-chloroform extracted, ethanol precipitated, and resolved on a 10% polyacrylamide-7M Urea gel. Gels were dried and exposed to a PhosphorImager, scanned using a Storm Scanner and quantitated using ImageQuant software. Uncoupled in vitro decay reactions were carried out with 32P-labeled mature single-stranded or double-stranded miRNAs (IDT) (Supplemental Table S2) and 50 μg of HEK293T cytoplasmic extract at 15°C. Reactions were stopped with ULB spiked with 1000 cpm 5’-end 32P-labeled 60-nt-long loading control.

**SUPPLEMENTAL MATERIAL**

Supplemental material can be found at http://www.rnajournal.org.

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